

# Absence of Plasmid-Mediated *mcr* Genes among Phenotypically Colistin-Resistant Carbapenem-Resistant Gram-Negative Isolates from a Tertiary Care Centre in Central India

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## ABSTRACT

**Background:** Colistin has re-emerged as a last-resort antimicrobial for treating infections caused by carbapenem-resistant Gram-negative bacteria (CR-GNB). The global discovery of plasmid-mediated mobilized colistin resistance (*mcr*) genes has raised serious concerns regarding the horizontal dissemination of colistin resistance. However, data on the molecular basis of colistin resistance in India remain limited.

**Objectives:** To detect plasmid-mediated *mcr-1* to *mcr-8* genes among phenotypically colistin-resistant carbapenem-resistant Gram-negative clinical isolates.

**Materials and Methods:** A total of 56 phenotypically colistin-resistant isolates identified by broth microdilution were subjected to polymerase chain reaction (PCR) for the detection of *mcr-1* to *mcr-8* genes using published primers and standardized protocols. Appropriate positive and negative controls were included.

**Results:** None of the 56 colistin-resistant isolates harbored *mcr-1* to *mcr-8* genes. These findings suggest that colistin resistance in the study isolates is predominantly mediated by chromosomal mechanisms rather than plasmid-borne resistance determinants.

**Conclusion:** The absence of *mcr* genes among phenotypically resistant isolates indicates a predominance of chromosomal resistance mechanisms in this region. Continuous molecular surveillance is essential to detect the future emergence of transferable colistin resistance and to inform antimicrobial stewardship strategies.

**Keywords:** Colistin resistance; *mcr* genes; Carbapenem-resistant organisms; PCR; Molecular epidemiology.

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## INTRODUCTION

Antimicrobial resistance (AMR) is a rapidly escalating global health crisis, threatening the effective management of bacterial infections. Among the most concerning developments is the rise of carbapenem-resistant Gram-negative bacteria (CR-GNB), including *Klebsiella pneumoniae*, *Acinetobacter baumannii*, and *Pseudomonas aeruginosa*, which are associated with high morbidity and mortality rates<sup>1</sup>.

The resurgence of colistin as a last-resort antibiotic has been driven by the limited availability of effective therapeutic alternatives for CR-GNB infections. Colistin exerts its bactericidal activity by binding to lipid A in the lipopolysaccharide (LPS) layer of Gram-negative bacteria, leading to membrane disruption and cell death<sup>2</sup>. Despite its effectiveness, increased clinical and agricultural use has resulted in the emergence of colistin resistance worldwide<sup>3</sup>.

A major breakthrough in understanding colistin resistance occurred in 2015 with the discovery of the plasmid-mediated *mcr-1* gene in *Escherichia coli* isolates from China<sup>4</sup>. Since then, multiple *mcr* variants (*mcr-1* to *mcr-10*) have been identified globally, raising concerns about horizontal gene transfer and rapid dissemination of resistance across bacterial species<sup>5</sup>. Unlike chromosomal mutations, plasmid-borne *mcr* genes pose a greater public health threat due to their mobility and potential for widespread transmission.

In India, reports of *mcr*-mediated resistance remain sporadic, and several studies have demonstrated colistin resistance in the absence of detectable *mcr* genes<sup>6–8</sup>. These observations suggest that chromosomal mechanisms, such as mutations in regulatory systems (*pmrAB*, *phoPQ*, *mgrB*) and lipid A modification pathways, may play a dominant role<sup>9</sup>. Given the clinical importance of distinguishing plasmid-mediated from chromosomal resistance, this study aimed to investigate the molecular basis of colistin resistance by detecting *mcr-1* to *mcr-8* genes among phenotypically colistin-resistant carbapenem-resistant isolates from a tertiary care hospital in central India.

## MATERIALS AND METHODS

### Study Design and Isolate Selection

This molecular analysis was conducted as part of a hospital-based cross-sectional study in the Department of Microbiology, Index Medical College Hospital & Research Centre, Indore, from January 2022 to June 2025.

A total of 56 non-duplicate Gram-negative bacterial isolates demonstrating phenotypic resistance to colistin (MIC  $\geq 4$   $\mu\text{g/mL}$  by broth microdilution) were included. These isolates were obtained from blood, urine, and pus samples of patients with clinically significant infections.

### Bacterial Identification and Phenotypic Testing

Initial identification and antimicrobial susceptibility testing were performed using the VITEK 2 Compact automated system (bioMérieux, France). Colistin susceptibility was confirmed using the broth microdilution method in accordance with CLSI 2023 guidelines<sup>10</sup>.

### DNA Extraction

Genomic DNA was extracted from overnight bacterial cultures using a commercially available extraction kit, following the manufacturer's protocol. The quality and quantity of extracted DNA were assessed spectrophotometrically.

### Polymerase Chain Reaction for *mcr* Genes

All isolates were screened for *mcr-1* to *mcr-8* genes using polymerase chain reaction (PCR). Published primer sequences and cycling conditions were employed for each target gene<sup>11</sup>. Each PCR run included known positive controls and nuclease-free water as a negative control.

Amplified products were analyzed by agarose gel electrophoresis and visualized under ultraviolet illumination after ethidium bromide staining.

## RESULTS

All 56 phenotypically colistin-resistant carbapenem-resistant Gram-negative isolates were subjected to molecular screening. None of the isolates showed amplification of *mcr-1*, *mcr-2*, *mcr-3*, *mcr-4*, *mcr-5*, *mcr-6*, *mcr-7*, or *mcr-8* genes.

The absence of plasmid-mediated *mcr* genes across isolates from diverse clinical specimens suggests that colistin resistance in this study population is not driven by transferable resistance determinants.

## DISCUSSION

The emergence of plasmid-mediated colistin resistance has generated global concern due to the potential for rapid horizontal dissemination of resistance genes across bacterial populations<sup>4,5</sup>. In this study, none of the phenotypically colistin-resistant isolates harbored *mcr* genes, indicating a predominance of chromosomal resistance mechanisms.

Similar findings have been reported from several Indian and international studies, where colistin resistance was observed in the absence of *mcr* genes<sup>6–8,12</sup>. Chromosomal mutations affecting regulatory pathways such as *pmrAB*, *phoPQ*, and *mgrB* have been implicated in lipid A modification, leading to reduced colistin binding<sup>13</sup>.

The absence of *mcr* genes in this study is reassuring from a public health perspective, as plasmid-mediated resistance poses a greater threat due to its mobility. However, chromosomal resistance remains clinically significant and may still lead to treatment failure if undetected<sup>14</sup>.

These findings underscore the importance of integrating molecular diagnostics with phenotypic testing. While routine *mcr* gene screening may not yet be warranted in all settings, periodic surveillance is essential to detect emerging plasmid-mediated resistance and to guide infection control policies<sup>15</sup>.

## CONCLUSION

This study demonstrates that phenotypic colistin resistance among carbapenem-resistant Gram-negative isolates in a tertiary care hospital in central India is not mediated by plasmid-borne *mcr-1* to *mcr-8* genes. The resistance is likely attributable to chromosomal mechanisms. Ongoing molecular surveillance, rational antimicrobial use, and robust stewardship programs are critical to preventing the emergence and spread of transferable colistin resistance.

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