Human Immune-Inflammatory Gene expression Profile after Infection with Human Papillomavirus

Amina Thannon Hamed AL-Dabbagh¹, Abdulrhem Thonnon Younis AL-Ghazal²
Department of Biology, College of Science, University of Mosul, Mosul City, Iraq

Abstract: Human papillomavirus (HPV) is one of the most common causes of sexually transmitted disease worldwide in both men and women. HPV is associated with a variety of diseases ranged from innocuous lesions to cancer. Three Data-groups of up regulated immune-inflammatory genes (25 genes with 3 replicates) that affected by human papilloma virus especially types 16, 18, 31, and 45 were subjected to this study. Bioinformatics is an emerging scientific discipline—used in this study to organize, analyze, and distribute biological information of genes affected by HPV. Data analysis of Real-time-reverse transcription PCR-array (RT-PCR-array) is the most sensitive and reliable method used to analyze the human gene expression affected by HPV. Gene ontology is a new bioinformatics method mainly provides typical batch annotation and gene analysis used to highlight the most important functions of up regulated genes after HPV infections. The immune-inflammatory gene expression profile for host affected with HPV was: 25 up regulated genes with 6.47 fold change (CCL18) to 79.22 (IL1RN).

Keywords: Human papilloma virus, Gene expression, Real-Time-PCR Array, Bioinformatics, Gene Ontology.

Introduction

Human papillomavirus (HPV) is one of the most common causes of sexually transmitted disease worldwide in both men and women and is thought to be the most common sexually transmitted viral disease. Papillomaviruses are ubiquitous and have been detected in a wide variety of animals as well as in humans and are specific for their respective hosts. More than 200 types of HPV have been recognized on the basis of DNA sequence data showing genomic differences (Burd, 2003).HPV is associated with a variety of clinical conditions that range from innocuous lesions to cancer. Most HPV infections can cause warts (plantar warts, common warts and flat warts). Genital HPV types infect primarily the cervix, vagina, vulva, penis and anus. Among the cancers attributable to high-risk HPV infection, cervical cancer has received the most attention. HPV-16, -18, - 31, -45 accounts for more than 90% of cervical carcinomas. Of these types, HPV-16 is the most often found, accounting for about half of the cervical cancer cases in the United States and Europe. In addition, high-risk HPV types have been related with other genital cancers, such as carcinoma of vagina, vulva, penis and anus, and their precancerous lesions. (Gomez and Santos, 2007) Bioinformatics is an emerging scientific discipline that uses information technology to organize, analyze, and distribute biological information in order to answer complex biological questions (Abd-Elsalam 2003). Microarray analysis and RT-qPCR are powerful methods to determine global profiles of genes in cells and tissues under a variety of complex biological conditions (Clewley, 2004). Development of microarrays facilitated the screening of viral pathogens from across broad viral families (See and Work, 2008; Penelope, et al., 2004). Data analysis of Real-time-reverse transcription PCR-array (RT-PCR-array) (SABioscience) is the most sensitive and reliable method for gene expression analysis (Montgomery and Daum 2009) which can be used to analyze the results of previous studies (Performed on the effect of HPV on human immune genes) through the determination of the altered genes and connect them with the new advanced technique(Gene Ontology) which provides the scientific researcher with new and important information mainly provides typical batch annotation and gene analysis to highlight the most relevant pathways associated with a given gene list (Huang et al., 2009). In addition Gene Ontology can determine the sit effect of these altered genes on the most important immune-inflammatory pathways in patients affected with microbial infections (David, 2008). The aim of this study is to determine thehuman immune-inflammatory gene expression profile after infection with human Papillomavirus especially cancerous types 16, 18, 31, and 45. In addition to specify the up regulated genes and their function during HPV infections

Materials and Methods

Gene-Data Collection

Three groups of up regulated immune-inflammatory genes were collected from previous studies after infection with cancerous human papilloma virus especiallytypes 16, 18, 31, and 45. The collected genes that their gene expression changes (with different fold change) were classified into three replicates and regarded as the Acute sample (Table 1).

Table 1: Fold change of gene expression and replicates of immune-inflammatory genes after HPV infections.

| Up regulated genes | | | Fold change of infected genes by HPV | | | | |
|--------------------|----------|--------|--------------------------------------|--------|--------|--|--|
| Gene No. | Position | Symbol | Group1 | Group2 | Group3 | | |
| 1 | A02 | BCL6 | 2.0 | 2.5 | 3.0 | | |
| 2 | A11 | CCL17 | 2.0 | 2.07 | 2.4 | | |
| 3 | A12 | CCL18 | 1.8 | 1.5 | 2.0 | | |
| 4 | B01 | CCL19 | 1.8 | 1.9 | 2.0 | | |
| 5 | B02 | CCL2 | 2 | 3.1 | 4.3 | | |
| 6 | B03 | CCL20 | 2.0 | 2.4 | 3.6 | | |
| 7 | B04 | CCL21 | 2.0 | 5.9 | 9.8 | | |
| 8 | B12 | CCL7 | 2.3 | 9.3 | 5.8 | | |
| 9 | C08 | CCR7 | 2.0 | 2.1 | 2.0 | | |
| 10 | C11 | CEBPB | 2.0 | 2.4 | 4.8 | | |
| 11 | D02 | CXCL1 | 3.7 | 7.9 | 12.1 | | |
| 12 | D04 | CXCL11 | 2.3 | 3.1 | 3.5 | | |
| 13 | D12 | CXCL9 | 1.9 | 2.0 | 2.6 | | |
| 14 | E02 | IFNA2 | 2.0 | 4.8 | 8.9 | | |
| 15 | E03 | IL-10 | 2.0 | 4.6 | 6.0 | | |
| 16 | E04 | IL10RA | 2.0 | 13.0 | 24 | | |
| 17 | E08 | IL17C | 1.6 | 2.0 | 2.1 | | |
| 18 | E09 | IL-1A | 2.0 | 2.1 | 5.8 | | |
| 19 | E10 | IL-1B | 1.5 | 2.0 | 3.6 | | |
| 20 | F04 | IL36G | 2.0 | 6.9 | 11.0 | | |
| 21 | F06 | IL1RN | 1.61 | 6.6 | 11.6 | | |
| 22 | F10 | IL-8 | 2.0 | 2.5 | 2.1 | | |
| 23 | G06 | MIF | 3.4 | 13.00 | 24.4 | | |
| 24 | G09 | TNF | 1.8 | 2.0 | 2.5 | | |
| 25 | G10 | TOLLIP | 2.8 | 3.3 | 4.5 | | |

The up regulated genes with more than one fold change were compared with control group (One foldchange) which has been taken from standard tables of previous studies (Al-Ghazal, 2010).

Calculation of fold change

The fold changes of genes samples were changed into threshold values (Ct) which represent the initial cycle at which the DNA sample was amplified using Real Time PCR-Array-DataAnalysis (Table 2).

Table 2: Fold changes values concluded from genes undergoing high expression after HPV infections.

| Gene | Group | Ct | Ct | Group | Ct | Ct | Group | Ct | Ct |
|--------|-------|---------|-------|-------|---------|-------|-------|---------|-------|
| name | 1 | Control | Acute | 2 | Control | Acute | 3 | Control | Acute |
| BCL6 | 2.0 | 30.5 | 27.8 | 2.5 | 27.7 | 21.8 | 3.0 | 24.6 | 29.1 |
| CCL17 | 2.0 | 30.5 | 27.8 | 2.07 | 30.5 | 27.8 | 2.4 | 32.7 | 29.7 |
| CCL18 | 1.8 | 31.2 | 28.6 | 1.5 | 31.2 | 28.9 | 2.0 | 30.5 | 27.8 |
| CCL19 | 1.8 | 31.2 | 28.6 | 1.9 | 27.4 | 21.9 | 2.0 | 30.5 | 27.8 |
| CCL2 | 2 | 30.5 | 27.8 | 3.1 | 25.0 | 19.8 | 4.3 | 24.3 | 29.3 |
| CCL20 | 2.0 | 30.5 | 27.8 | 2.4 | 32.7 | 29.7 | 3.6 | 29.1 | 32.4 |
| CCL21 | 2.0 | 30.5 | 27.8 | 5.9 | 33.2 | 29.0 | 9.8 | 24.4 | 31.0 |
| CCL7 | 2.3 | 27.7 | 23.6 | 9.3 | 24.1 | 30.4 | 5.8 | 29.1 | 33.3 |
| CCR7 | 2.0 | 30.5 | 27.8 | 2.1 | 28.8 | 24.4 | 2.0 | 30.5 | 27.8 |
| CEBPB | 2.0 | 30.5 | 27.8 | 2.4 | 32.7 | 29.7 | 4.8 | 23.4 | 28.9 |
| CXCL1 | 3.7 | 31.5 | 27.9 | 7.9 | 28.0 | 21.5 | 12.1 | 22.0 | 27.0 |
| CXCL1 | 2.3 | 27.5 | 23.6 | 3.1 | 25.0 | 19.9 | 3.5 | 32.7 | 29.2 |
| 1 | | | | | | | | | |
| CXCL9 | 1.9 | 27.4 | 21.9 | 2.0 | 30.5 | 27.8 | 2.6 | 26.6 | 30.9 |
| IFNA2 | 2.0 | 30.5 | 27.8 | 4.8 | 28.9 | 23.4 | 8.9 | 23.2 | 27.6 |
| IL-10 | 2.0 | 30.5 | 27.8 | 4.6 | 27 | 21.3 | 6.0 | 23.7 | 29.6 |
| IL10RA | 2.0 | 30.5 | 27.8 | 13.0 | 33.6 | 28.2 | 24 | 24.0 | 29.0 |
| IL17C | 1.6 | 29.6 | 24.3 | 2.0 | 30.5 | 27.8 | 2.1 | 24.4 | 28.8 |
| IL-1A | 2.0 | 30.5 | 27.8 | 2.1 | 28.8 | 24.0 | 5.8 | 29.1 | 33.3 |
| IL-1B | 1.5 | 31.2 | 28.9 | 2.0 | 30.5 | 27.8 | 3.6 | 29.1 | 32.4 |

| IL36G | 2.0 | 30.5 | 27.8 | 6.9 | 31.3 | 25.7 | 11.0 | 25.0 | 31.3 |
|--------|-----|------|------|------|------|------|------|------|------|
| IL1RN | 1.6 | 29.6 | 24.3 | 6.6 | 31.3 | 25.7 | 11.6 | 20.6 | 27.7 |
| IL-8 | 2.0 | 30.5 | 27.8 | 2.5 | 27.7 | 21.8 | 2.1 | 24.4 | 28.8 |
| MIF | 3.4 | 28.2 | 32.7 | 13.0 | 33.6 | 28.2 | 24.4 | 24.5 | 29.3 |
| TNF | 1.8 | 31.2 | 28.6 | 2.0 | 30.5 | 27.8 | 2.5 | 21.8 | 27.7 |
| TOLLIP | 2.8 | 24.6 | 21.7 | 3.3 | 23.3 | 26.4 | 4.5 | 24.2 | 29.2 |

Analysis of CT values

The Ct values introduced into gene analysis site (SABioscience-RT-PCR-Array using a Blank Excel Spread-sheet, SABioscience Data Analysis Template Excel File (Campeau et al., 2009; Myskiw et al., 2009). The results of this analysis represent the new fold change values (3 replicas) of gene expression compared with the same control group in all genetic analysis (Table 2). After gene analysis the value of gene expression for each gene was determined; more than one fold (compared with one fold for control genes) regarded as up regulated gene. Those with less than one fold regarded as down regulated gene.

Genes of RT-PCR-Array

The up regulated genes collected from previous studies will introduced in SABioscience-RT-PCR-Array technique. Using a Blank Excel Spread-sheet SABioscience Data Analysis Template Excel File which includes the following 84 immune genes: Cytokine genes, Cytokine receptors genes ,Chemokine receptors and other genes involved in inflammatory response.

Gene Ontology

For gene ontology analysisthe gene code number (Unigene or Refseq), of up regulated genes (Table-2), will interred in the data analysis using: David Functional Annotation Bioinformatics Microarray Analysis, Gene-annotation enrichment analysis, Functional annotation clustering, KEGG pathway mapping, and gene-disease association.

Statistical analysis

Since each gene has 6 values (3 values for acute samples and 3 values for control samples). All control values of all genes tested against all acute values and the differences between the two values were analyzed using the K- paired sample (Friedman test) to calculate significance degree between control and acute (P-value)

Result and Discussion

The results showed that HPV especially the cancerous types cause different changes in the expression of 25 human immune-inflammatory genes in different studies (p<0.021). These changes classified into three groups (Table 2). First group inclued genes scattered randomly with different values ranged between 6.4 fold change in IL1B to 59.7 fold changeinCXCL9 gene (figure 1). This means that HPV cause moderate impact on host human cell and stimulate gradual expression of 25 different human immune-inflammatory genes (figure 2).

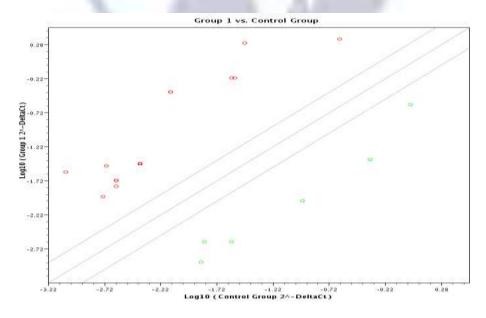


Figure 1: Scattered plot: Up regulated (Red circles) and down regulated (Green circles) genes in Acute 1 (Some circles may express more than one gene with same fold change).

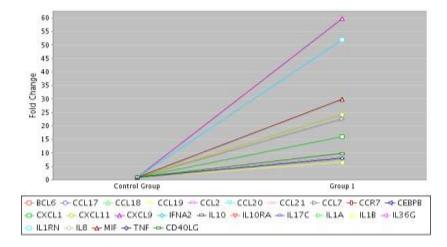


Figure 2: Multi curve: Up regulated genes (25 genes) for Acute 1

The second group of up regulated genes showed higher expression of genes ranged between 6.4 fold changes in 4 genes (TNF, IL1B,IL17C, and CXCL9) and 90.5 fold changes in CXCL1 gene (Figure 3). These results gave indicator that the effect of HPV on gene expression may vary according to the different types of HPV and to the immune status for different patients subjected to different studies (Figure 4).

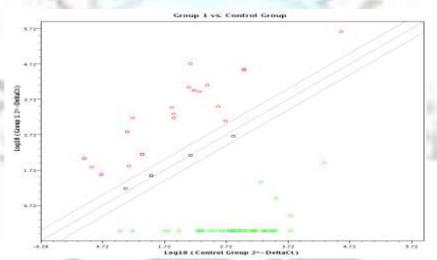


Figure 3: Gene analysis of Acute group 2 with 25 affected genes after HPV infections

Scattered plot: Up regulated (Red circles) and down regulated (Green circles) genes in Acute 2 (Some circles may express more than one gene (genes with same fold change).

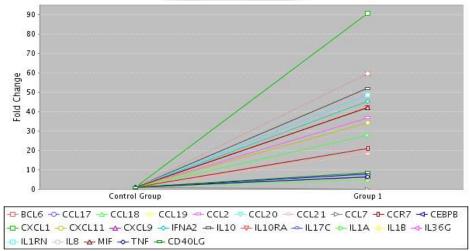


Figure 4: Multi curve: Up regulated genes (25 genes) for Acute 2 compared with control

The highest effect of HPV on the expression of human immune-inflammatory genes was revealed in the third group (Figure 5). The expression ranged between 6.4 fold change in CCL18 and 137.187 in IL1RN which represents the most affected gene after infection with HPV (Figure 6).

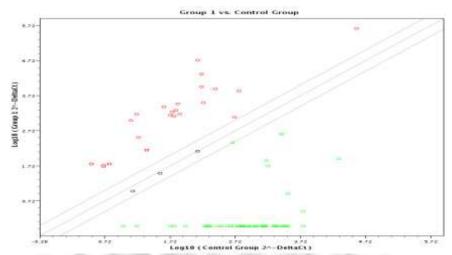


Figure 5: Gene analysis of Acute group 3 with 25 affected genes after HPV infections. Scattered plot: Up regulated (Red circles) and down regulated (Green circles) genes in Acute 3 (Some circles may express more than one gene (genes with same fold change.

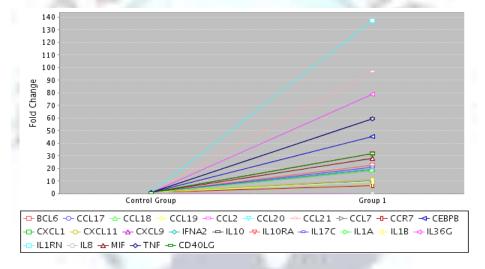


Figure 6: Multi curve: Up regulated genes (25 genes), for Acute 3.

The results showed that the most common affected genes were 11, those with mean of 22.77 to 79.22 fold changes in their expression (Table 3).

Table 3: The most important up regulated genes (22.77to 79.22 fold change) affected after HPV infection

| | G | | | | Fold |
|----|--------|---------|---------|---------|--------|
| | Gene | Group1 | Group2 | Group3 | change |
| | symbol | • | • | • | (mean) |
| 1 | IL1RN | 51.9842 | 48.5029 | 137.187 | 79.22 |
| 2 | CXCL1 | 16 | 90.5097 | 32 | 46.16 |
| 3 | IL36G | 8.5742 | 48.5029 | 78.7932 | 45.28 |
| 4 | CCL21 | 8.5742 | 18.3792 | 97.0059 | 41.31 |
| 5 | IL10 | 8.5742 | 51.9842 | 59.7141 | 40.08 |
| 6 | MIF | 29.8571 | 42.2243 | 27.8576 | 33.30 |
| 7 | BCL6 | 8.5742 | 59.7141 | 22.6274 | 30.3 |
| 8 | CXCL9 | 59.7141 | 6.498 | 19.6983 | 28.63 |
| 9 | IL10RA | 8.5742 | 42.2243 | 32 | 27.59 |
| 10 | IL17C | 51.9842 | 6.498 | 21.1121 | 26.52 |
| 11 | IFNA2 | 8.5742 | 45.2548 | 19.6983 | 22.77 |

International Journal of Enhanced Research in Science Technology & Engineering, ISSN: 2319-7463 Vol. 3 Issue 5, May-2014, pp: (324-331), Impact Factor: 1.252, Available online at: www.erpublications.com

Bioinformatics analysis using David Functional Annotation Bioinformatics Microarray Analysis, revealed that these up regulated 11 genes by HPV were mediated important inflammatory functions in human represented by: immune system processes, response to stimuli, locomotion, multi-organism process, developmental process, biological regulation.Intracellular Interleukin-1 Receptor Antagonist (IL1RN) was increased with 79.22 fold change which is more than other study (Chevalier et al.,2013) which also showed increasing in the fold change. Over expression of this gene is very important because it stimulates production of protein which inhibits the activities of interleukin 1, alpha (IL1A) and interleukin 1, beta (IL1B), and modulates a variety of interleukin 1 related immune and inflammatory responses (Szaboet al., 2010).

The mRNA transcript of Chemokine (C-X-C motif) ligand 1 (CXCL1) produced with 46.16 fold after HPV which regarded as a high level compared with other study (DeVotiet al., 2008) which showed 3.1 fold change only. Up regulation of this gene reflected its immunological roles such as stimulation the expression of beta-defensin which leads to antimicrobial barrier formation. In addition acts as a chemoattractant for neutrophils (Nischalkeet al., 2012). Interleukine-36 gamma gene (IL36G) (also called IL-1F9)increased with 45.28 fold change whereas in other study increased up to 13.1fold change (DeVotiet al 2008). This increasing was stimulated by HPV infection and immune system activates signaling transduction chainto activateslocal inflammatory response against HPV infections (Taylor et al., 2002). The expression of this cytokine in keratinocytes can also be induced by a contact hypersensitivity reaction or viral infection (Bachmann et al., 2012)

Chemokine ligand 21 (CCL21) increased with 41.31 fold change whereas in other study it was also increased but in low limit (more than 2 fold) (Trimble et al. 2010). This high increasing of this Chemokine is associated with, Inhibition of hemopoiesis, stimulation of chemotaxisthymocytes and activation of T-cells (Oliveira et al., 2013).. Interlukine-10 (IL-10) increased with 40.08 fold changes which in turn enhances B cell survival, proliferation, and antibody production (Zhenget al., 2012; Rezaeiet al., 2010). Many other studies also revealed that IL10 gene expression increased highly during infection and this high level of this protein enhance viral survive (Bermdez-Morales et al., 2011; Torres-Povedaet al., 2012; AL-Ghazal, 2010). The infection with HPV stimulates the host cells to activate expression of MIF gene (33.30 fold). In other study this gene also up regulated (14.352 fold change) (Savolaet al., 2011). The expression of MIF at sites of inflammation suggests a role as mediator in regulating thefunction of macrophages in host defense (Assuncaoet al., 2010).BCL6 gene ispro-inflammatory cytokine up regulated with 30.03 fold change in this study. This gene stimulates innate immunity against microbial infections and produced in inflammatory sites mediating the regulation of macrophage functions(Lo et al., 2013).

The mRNA of CXCL9 gene expressed with 28.63fold change after HPV infection. Other study revealed more upregulation (79 fold change) (AL-Ghazal, 2010). This increasing reflexes its important functions through stimulation of immune response and recruits T-cells to inflammatory sites (Lau and Peiris, 2009). The next up regulated gene in this study was Intrleukine-10 receptor alpha (IL10RA) with 27.59 Fold change which came in agreement with other study (24 fold change)(Baumgarthet al., 2004). IL10RA play key role in stimulation of high level of IL10 expression in monocytes, B-cells, large granular lymphocytes and T-cells. It has been shown to mediate the immunosuppressive signal of interleukin 10, and thus inhibits the synthesis of proinflammatory cytokines(Huangeet al., 2009). The expression of IL17C plays key role in innate immunity, this gene wasupregulated with 26.52 fold change. It produces peptides act as proinflammatory molecules against microbes through stimulation of NF-kappa-B and MAPK pathways. Other study also revealed increasing in IL17C level during microbial infections (Pfeifer et al., 2013). The mRNA of Interferon alpha 2 (IFNA2) gene up regulated with 24.50 fold change whereas in previous studies showed lower production of IFNA2 represented by 17.3 fold change (AL-Ghazal 2012) and 12.67 fold change (Ubolet al., 2008). IFNA2 induces hundreds of IFN stimulating genes (ISGs) which activate antiviral state in the infected cells. This protein also has been shown to be effective in reducing the symptoms and duration of the viral infections (Haoet al., 2013)

Conclusion

In conclusion the human immune system has impact factor on HPV through the up regulation of 25 key inflammatory genes. Eleven of these genes were highly produced compared with others. The immune-inflammatory gene expression profile for human patients affected with HPV was: 25 up regulated genes with mean of upregulation ranged between 6.47fold change for CCL18and 79.22 for IL1RN.

References

[1]. Abd-Elsalam, A. Kamel (2003). Bioinformatics tools and guideline for PCR primer design, African Journal of biotechnology, Vol. 2(5), pp 91 – 95.

International Journal of Enhanced Research in Science Technology & Engineering, ISSN: 2319-7463 Vol. 3 Issue 5, May-2014, pp: (324-331), Impact Factor: 1.252, Available online at: www.erpublications.com

- [2]. AL-Ghazal, A. T. (2010). Effect of viral respiratory infections by influenza A virus, Parainfluenza viruses and Adenoviruses on Immuno-inflammatory gene expression /inhibitory of human white blood cells.Ph.D thesis. University of Jordan, Amman, Jordan, 179-190.
- [3]. Al-Ghazal A.T.Y. (2012). Determination of the Changes in Human Leukocytes Inflammatory Genes Expression after Infection with Human Papillomavirus, Raf. J. Sci., 23, No.4 pp 33-50, 2012.
- [4]. Assuncao-Miranda, Bozza MT, Da Poian AT (2010) Pro-inflammatory response resulting from sindbis virus infection of human macrophages: implications for the pathogenesis of viral arthritis, J Med Virol. 82(1):164-74.
- [5]. Bachmann M, Scheiermann P, Hardle L, Pfeilschifter J, Mühl H. (2012) IL-36γ/IL-1F9, an innate T-bet target in myeloid cells. J Biol Chem.287(50):41684-96.
- [6]. Baumgarth Nicole, Richard Szubin, Greg M. Dolganov, Mitchell R. Watnik, Deborah Greenspan, Maria Da Costa, Joel M. Palefsky, Richard Jordan, Mario Roederer, and John S. Greenspan (2004) Highly Tissue Substructure-Specific Effects of Human Papilloma Virus in Mucosa of HIV-Infected Patients Revealed by Laser-Dissection Microscopy-Assisted Gene Expression Profiling. American Journal of Pathology, 165, No. 3,
- [7]. Bermudez-Morales, V.H., Peralta-Zaragoza, O Alcocer Gonzalez.J.M., Moreno J.and Madrid- V. (2011) Marina IL-10 expression is regulated by HPV E2 protein in cervical cancer cells. Molecular Medicine REPORTS 4: 369-375.
- [8]. Burd Eileen M. (2003). Human papillomavirus and cervical cancer, Clinical Microbiology Review ,16, No. 1, p(1-17).
- [9]. Campeau, P., Rafei, M., Boivin, M., Sun, Y., Grabowski, G., Galipeau, J. (2009). Characterization of Gaucher disease bone marrow mesenchymal stromal cells reveals an altered inflammatory secretome, Blood, 114(15). 3181-3190.
- [10]. Clewley J.(2004) Arole of arrays in clinical virology: fact or fiction, Journal of Clinical Virology, 29, 2-12.
- [11]. David , F., Farley ,J., Huang ,H., Lavoie, J., Laverty, S., (2008) .cytokine and chemokine gene expression of IL-1 beta stimulated equine articular chondrocyte, Veterinary Surgery . 37(5) .499.
- [12]. DeVoti James A, David W Rosenthal, Rong Wu, Allan L Abramson, Bettie M Steinberg, and Vincent R Bonagura (2008) Immune Dysregulation and Tumor-Associated Gene Changes in Recurrent Respiratory Papillomatosis: A Paired Microarray Analysis, M O L M E D 1 4 (9 10) 6 0 8 6 17.
- [13]. Gomez Daniel tena and Santos Juan López,(2007).human papillomavirus infection and cervical cancer: pathogenesis and epidermology, Communication Current Research And Eeducation Topics And Trends In Applied Microbiology AMendez Vilas, 2 (680-688).
- [14]. Hao J Jin W, Li X, Wang S, Zhang X, Fan H, Li C, Chen L, Gao B, Liu G, Meng S. (2013) Inhibition of alpha interferon (IFN-α)-induced microRNA-122 negatively affects the anti-hepatitis B virus efficiency of IFN-α. J Virol.87 (1):137-47.
- [15]. Huang D, Sherman B, Limpicki R. (2009).systematic and integrative analysis of large genes lists using DAVID Bioinformatics Resources, Nature Protocol, 4(1).44-57
- [16]. Lau Y.L. and Peiris J.S. (2009). Association of cytokine and chemokine gene polymorphisms with severe acute respiratory syndrome, 15 suppl.2;43-6.
- [17]. Lo MC, Yip TC, Nagan KC, Cheng WW, Law CK, Chan PS, Chan KC, Wong CK, Wong RN, Lo KW, NG WT, Lee WM, Tsao SW, Kwong LW, Lung ML, Mak NK.. (2013) Role of MIF/CXCL8/CXCR2 signaling in the growth of nasopharyngeal carcinoma tumor spheres. 10;335(1):81-92
- [18]. Montgomery C, Daum R., (2009) transcription of inflammatory genes in the lung after infection, Infectious immunology, 77(5). 2159-67.
- [19]. Myskiw C, Arsenio J, van Bruggen R, Deschambault Y, Cao J. (2009). Vaccinia virus E3 suppresses expression of diverse cytokines through inhibition. Journal of Virology, 83(13):6757-68.
- [20]. Nischalke HD, Berger C, Luda C, Müller T, Berg T, Coenen M, Kramer B, Korner C, Trebicka J, Grunhage F, Lammert F, Nattermann J, Sauerbruch T, Spengler U. (2012) The CXCL1 rs4074 A allele is associated with enhanced CXCL1 responses to TLR2 ligands and predisposes to cirrhosis in HCV genotype 1-infected Caucasian patients. J epatol; 56(4):758-64.
- [21]. Pfeifer P, Voss M, Wonnenberg B, Hellberg J, Seiler F, Lepper PM et al., (2013) IL-17C is a mediator of respiratory epithelial innate immune response. Am J Respir Cell MolBiol; 48(4):415-21.
- [22]. Oliveira-Neto HH1, de Souza PP, da Silva MR, Mendonça EF, Silva TA, Batista AC. (2013). The expression of chemokines CCL19, CCL21 and their receptor CCR7 in oral squamous cell carcinoma and its relevance to cervical lymph node metastasis. Tumour Biol. 34(1):65-70.
- [23]. Penelope A., Bryant D., Roy R., and Nigel C., (2004). Chips with every Thing: DNA microarray infectious diseases, Lancet Infectious Diseases, 4, 100-111.
- [24]. Rezaei N, Aghamohammadi A, Mahmoudi M, Shakiba Y, Kardar GA, Mahmoudi M, Moradi B, Amirzargar AA. (2010) Association of IL-4 and IL-10 gene promoter polymorphisms with common variable immunodeficiency. Immunology ;215(1):81-7.
- [25]. SavolaSuvi, ArtoKlami, SamuelMyllykangas, CristinaManara, Katia Scotlandi, PieroPicci, SakariKnuutila, and JukkaVakkila, (2011)High Expression of Complement Component 5 (C5) at Tumor Site Associates with Superior Survival in Ewing's Sarcoma Family of Tumour Patients, ISRN Oncology, Article ID 16, 87, pages 12-10.
- [26]. See H., and Work p., (2008). Innate immune response to viral infection of lungs , Paediatric Respiratory Reviews , 9,243-250.

International Journal of Enhanced Research in Science Technology & Engineering, ISSN: 2319-7463 Vol. 3 Issue 5, May-2014, pp: (324-331), Impact Factor: 1.252, Available online at: www.erpublications.com

- [27]. Szabo K Olofsson, Sheikine Y, Jatta K, Ghaderi M, Samnegard A, Eriksson P, Sirsjo A. (2009). A functional interleukin-1 receptor antagonist polymorphism influences atherosclerosis development. The interleukin-1beta:interleukin-1 receptor antagonist balance in atherosclerosis .Circ J.73(8):1531-6..
- [28]. Taylor SL, Renshaw BR, Garka KE, Smith DE, Sims JE.(2002) Genomic organization of the interleukin-1 locus. Genomics; 79(5):726-33.
- [29]. Torres-Poveda, Kirvis, Ana I Burguete-Garcia1, Miguel Cruz, Gabriela A Martinez-Nava, Margarita Bahena-Roman, Esmeralda Ortiz-Flores, Abrahan Ramirez-Gonzalez, Guillermina Lopez-Estrada, Karina Delgado-Romero and Vicente Madrid-Marina. (2012) The SNP at -592 of human IL-10 gene is associated with serum IL-10 levels and increased risk for human papillomavirus cervical lesion development. Infectious Agents and Cancer 2012, 7:32
- [30]. Trimble Cornelia L, and Achim A. JungbluthFerdynand Kos, Jessica Teague, Ying Jiang, Nicole C. Barat Thoburn, Nicole C. Hanson, Jodie Tassello, Denise Frosina, Trimble, Rachael A. Clark, Christopher (2010) Epithelium Excludes CD8 T Cells from Dysplastic Cervical Intraepithelial Neoplasia in Humans Human Papillomavirus 16-Associated. J Immunol; 185:7107-7114;
- [31]. UbolSukathida, PromsinMasrinoul, JeerayutChaijaruwanich, SiripenKalayanarooj, TakolCharoensirisuthikul, and JitraKasisith(2008) Differences in Global Gene Expression in Peripheral Blood Mononuclear Cells Indicate a Significant Role of the Innate Responses in Progression of Dengue Fever but Not Dengue Hemorrhagic Fever. The Journal of Infectious Diseases, 197:1459–67.
- [32]. Zheng Bi, X, T, Lan Q, Xu Z, Chen Y, Zhu G, Foss F, Kim C, Dai M, Zhao P, Holford T, Leaderer B, Boyle P, Deng Q, Chanock SJ, Rothman N, Zhang Y.(2012) Genetic polymorphisms in IL10RA and TNF modify the association between blood transfusion and risk of non-Hodgkin lymphoma. Am J Hematol.87(8):766-9.