

Quantitative and qualitative analysis of the Human Microbiota in Health and Disease: A Review

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ABSTRACT

Microbiota has evolved with and continue to live on and within human beings. Environmental factors can affect intestinal microbial imbalance, which has a close relationship with human health and disease. Here, we focus on the interactions between the human microbiota and the host in order to provide an overview of the microbial role in basic biological processes and in the develop- ment and progression of major human diseases such as infectious diseases, liver diseases, gastrointesti- nal cancers, metabolic diseases, respiratory diseases, mental or psychological diseases, and autoimmune diseases. We also review important advances in techniques associated with microbial research, such as DNA sequencing, metabonomics, and proteomics combined with computation-based bioinformatics. Current research on the human microbiota has become much more sophisticated and more comprehen- sive. Therefore, we propose that research should focus on the host-microbe interaction and on cause- effect mechanisms, which could pave the way to an understanding of the role of gut microbiota in health and disease, and provide new therapeutic targets and treatment approaches in clinical practice.

Keywords: Microbiome Health Infectious disease Liver diseases, Gastrointestinal malignancy Metabolic disorder Microbiota technology Probiotics

INTRODUCTION

The human microbiota affects host physiology to a great extent. Over time, host-bacterial associations have developed into beneficial relationships. Symbiotic bacteria metabolize indigestible compounds, supply essential nutrients, defend against colonization by opportunistic pathogens, and contribute to the formation of intestinal architecture The human microbiota, especially the gut microbiota, has even been considered to be an "essential organ" [1], carrying approximately 150 times more genes than are found in the entire human genome [2]. Important advances have shown that the gut microbiota is involved in basic human biological processes, including modulating the metabolic phenotype, regulating epitheli- al development, and influencing innate immunity [3–6]. Chronic diseases such as obesity, inflammatory bowel disease (IBD), di- abetes mellitus, metabolic syndrome, atherosclerosis, alcoholic liver disease (ALD), nonalcoholic fatty liver disease (NAFLD), cir- rhosis, and hepatocellular carcinoma have been associated with the human microbiota [7,8] (Fig.1).

In recent decades, a tremendous amount of evidence has strong- ly suggested a crucial role of the human microbiota in human health and disease [7,9–23] via several mechanisms. First, the microbiota has the potential to increase energy extraction from food [24], increase nutrient harvest [9,10], and alter appetite signaling [25,26]. The microbiota contains far more versatile metabolic genes than are found in the human genome, and pro- vides humans with unique and specific enzymes and biochemical pathways [9]. In addition, a large proportion of the metabolic microbiotic processes that are beneficial to the host are involved in either nutrient acquisition or xenobiotic processing, including the metabolism of undigested carbohydrates and the biosynthe- sis of vitamins [10]. Second, the human microbiota also provides a physical barrier, protecting its host against foreign pathogens through competitive exclusion and the production of antimicro- bial substances [11–13]. Finally, the microbiota is essential in the development of the intestinal mucosa and immune system of the host [14,16]. For example, germ-free (GF) animals have abnor- mal numbers of several immune cell types, deficits in local and systemic lymphoid structures, poorly formed spleens and lymph





Fig. 1. Human microbial symbiosis has a close relationship with diseases of differ- ent systems.

nodes, and perturbed cytokine levels [16]. Studies on GF animals have suggested that the immune modulation functions of the microbiota are primarily involved in promoting the maturation of immune cells and the normal development of immune functions [14]. In addition, studies have revealed the central role of micro- bial symbiosis in the development of many diseases [17], such as infection [18], liver diseases [19], gastrointestinal (GI) malignancy [20], metabolic disorders [7], respiratory diseases [21], mental or psychological diseases [22], and autoimmune diseases [23].

In this article, we provide an overview of the role of the human microbiota in health and disease, the advent of microbiome-wide association studies, and potential and important advances in the development of clinical applications to prevent and treat human disease.

The human microbiota in health

The human microbiota affects host physiology to a great ex- tent. Trillions of microbes colonize the human body, including bacteria, archaea, viruses, and eukaryotic microbes. The body contains at least 1000 different species of known bacteria and carries 150 times more microbial genes than are found in the entire human genome [2]. Microbiotic composition and function differ according to different locations, ages, sexes, races, and diets of the host [27].

Commensal bacteria colonize the host shortly after birth. This simple community gradually develops into a highly diverse eco- system during host growth [28]. Over time, host-bacterial asso- ciations have developed into beneficial relationships. Symbiotic bacteria metabolize indigestible compounds, supply essential nu- trients, defend against colonization by opportunistic pathogens, and contribute to the formation of intestinal architecture [29]. For example, the intestinal microbiota is involved in the digestion of certain foods that cannot be digested by the stomach and small intestine, and plays a key role in maintaining energy homeosta- sis. These foods are primarily dietary fibers such as xyloglucans, which are commonly found in vegetables and can be digested by a specific species of Bacteroides [30]. Other non-digestible fibers, such as fructooligosaccharides and oligosaccharides, can be uti-lized by beneficial microbes, such as Lactobacillus and Bifidobac- terium [31]. Studies have clarified the role of the gut microbiota in lipid and protein homeostasis as well as in the microbial syn- thesis of essential nutrient vitamins [32]. The normal gut micro- biome produces 50-100 mmol·L⁻¹ per day of short-chain fatty acids (SCFAs), such as acetic, propionic, and butyric acids, and serves as an energy source to the host intestinal epithelium [33]. These SC- FAs can be quickly absorbed in the colon and serve many diverse roles in regulating gut motility, inflammation, glucose homeostasis, and energy harvesting [34,35]. Furthermore, the gut microbiota has been shown to deliver vitamins to the host, such as folates, vita- min K, biotin, riboflavin (B2), cobalamin (B12), and possibly other B vitamins. A previous study demonstrated that B₁₂ can be produced from delta-aminolevulinate (ALA) as a precursor [36].

In addition, gut-colonizing bacteria stimulate the normal de- velopment of the humoral and cellular mucosal immune systems [37]. The signals and metabolites of microorganisms can be sensed by the hematopoietic and non-hematopoietic cells of the innate immune system and translated into



physiological responses [38]. Studies comparing normal mice with GF mice have found that GF mice show extensive defects in the development of gut-associated lymphoid tissue and antibody production [29,39]. A report has also demonstrated that the gut microbiota generates a tolerogenic response that acts on gut dendritic cells and inhibits the type 17 T-helper cell (Th17) anti-inflammatory pathway [40]. However, notall microbiota lead to health benefits. Some induce inflammation under certain conditions.

The human microbiota in disease

The human microbiota and infectious diseases

Infection is one of the most common diseases caused by dys- biosis of the microbiota. Importantly, infectious disease and its treatment have a profound impact on the human microbiota, which in turn determines the outcome of the infectious disease in the human host (Fig. 2). Offending pathogens colonize the intesti- nal mucosa, thus resulting in the induction of a strong inflamma- tory response, followed by the translocation of the intestinal bac- teria [41,42]. Numerous studies have demonstrated the intimate relationship between infection and dysbiosis of the microbiota, and have shown that infection is associated not only with the microbiome, but also with viruses [43,44]. For example, the intes- tinal microbiota of patients with *Clostridium difficile* (*C. difficile*) infection (CDI) is significantly altered [45,46]. Disturbance of the microbiota is also associated with the progression of human immunodeficiency virus (HIV) [44,47], hepatitis B virus (HBV) [48], and other diseases [49,50].

Infection with Clostridium difficile

The pathological overgrowth of *C. difficile* is usually related to antibiotic-associated diarrhea, which is one of the most frequent complications following antibiotic administration and which is now a growing public health threat [45]. *C. difficile* is an anaero- bic, gram-positive, spore-forming bacillus that is a component of the human gut microbiota. Antibiotics disturb intestinal mucosa homeostasis, thus decreasing resistance against toxin-producing

C. difficile and promoting the progression of CDI [45]. Gu et al. [45] found that fecal bacterial diversity is reduced and the microbial composition dramatically shifts in patients following antibiotic administration, whether or not CDI is present. A decrease in putative butyrate-producing anaerobic bacteria and an increase in endo- toxin-producing opportunistic pathogens and lactate-producing phylotypes have been detected in patients following antibiotic administration, whether or not CDI is present [45]. Putative butyrate-producing anaerobic bacteria are significantly depleted

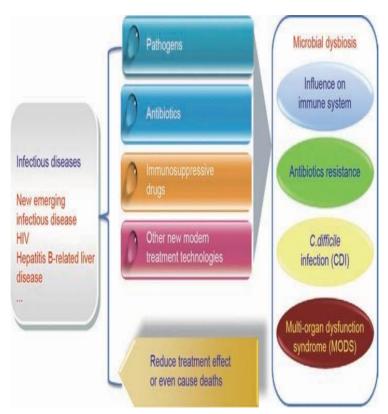


Fig. 2. Infectious diseases have a profound impact on the human microbiota. The wide use of antibiotics, immunosuppressive drugs, and other new treatment technologies for infectious diseases such as frequently emerging infectious diseases, HIV infection, and CDI has a profound



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in patients with antibiotic treatment when compared with healthy controls. The above changes in microbial communities may in- crease susceptibility to *C. difficile* colonization. Ling et al. [46] found that different toxigenic *C. difficile* strains have different effects on fecal microbiota in children. *C. difficile* strains that are both toxin A-positive and toxin B-positive reduce fecal bacteria diversity to a greater degree than strains that are only toxin B-positive.

Infection with Helicobacter pylori

Helicobacter pylori (H. pylori) is a pathogen that induces pep- tic disease. It was recently found to be related to the progress of periodontitis [49]. Hu et al. [49] investigated the correlation of H. pylori infection with periodontal parameters, periodontal path- ogens, and inflammation. Their study showed that the frequen- cies of Porphyromonas gingivalis, Prevotella intermedia, Fusobacte- rium nucleatum, and Treponema denticola are significantly higher in patients infected with H. pylori than in those without infection, whereas the frequency of Aggregatibacter actinomycetemcomitans is lower. The results indicate that patients with H. pylori show sig- nificantly higher probing depth and attachment loss, and that H. pylori might promote the growth of some periodontal pathogens and aggravate the progress of chronic periodontitis [49].

Bacterial vaginosis

Another important infection called bacterial vaginosis (BV) is associated with numerous adverse health outcomes including pre-term birth and the acquisition of sexually transmitted infec- tions. BV is regarded as an ecological disorder of the vaginal mi- crobiota. Using culture-independent polymerase chain reaction (PCR) denaturing gradient gel electrophoresis (DGGE) and bar- coded 454 pyrosequencing methods, Ling et al. [50] observed a profound shift in the absolute and relative abundances of bacte- rial species present in the vagina. In a comparison of populations associated with healthy and diseased conditions, three phyla and eight genera were clearly and strongly associated with BV. These genera may be used as targets for clinical BV diagnosis by means of molecular approaches [50].

Infection with HIV

At present, HIV continues to be a major global public health issue. The gut microbiomes in patients with HIV are significantly disturbed, and there are significant increases in the Firmicutes/ Bacteroidetes ratio of patients infected with HIV-1 [47]. Although the viral loads of HIV-1 are reduced after a short-term course of effective highly active anti-retroviral therapy (HAART), the diver- sity and composition of the fecal microbiota are not completely restored, and the dysbiosis remains [47]. South African teenage girls and young women have extremely high rates of HIV infec- tion, a phenomenon that has been considered to be associated with biological factors. Recently, one study found that the vaginal microbiome might affect the risk of HIV infection. A bacterium in the vagina named *Prevotella bivia* has been identified as causing inflammation. A microbicidal gel is available that decomposes the anti-HIV drug tenofovir, thus leading to tenofovir treatment fail- ure [44]. Through close examination of the vaginal microbiome, Cohen [44] recently found an unusual bacterium named *Gard- nerella* in the vagina; this finding potentially explains the high infection rates in South African women and strongly suggests that the vaginal microbiome affects HIV risk. Cohen [44] found that *Gardnerella* "gobbles up" tenofovir, thus rapidly decreasing the levels of the drug and leading to tenofovir treatment failure.

The human microbiota and liver diseases

Growing evidence demonstrates the close interaction of the GI tract (GIT) and the liver, as well as the chronic exposure of the liver to gut-derived factors including bacteria and bacterial components, thus fostering the use of the term "gut-liver axis" [51]. The intestinal microbiota produces ethanol, ammonia, and acet-aldehyde; these products may influence liver function through endotoxin release or liver metabolism [52].

Alterations in the intestinal microbiota play an important role in inducing and promoting liver damage progression as well as in direct injury resulting from different causal agents (e.g., viral, toxic, and metabolic agents) [53] through mechanisms such as the activation of Kupffer cells by bacterial endotoxins. The gut microbiota participates in the pathogenesis of liver cirrhosis complications, such as infections, spontaneous bacterial peri- tonitis, hepatic encephalopathy, and renal failure. Patients with liver cirrhosis have an altered bacterial composition in their gut; patients in Child-Pugh classes B/C have a higher prevalence of bacterial overgrowth than those in class A [54,55]. Fecal microbi- al communities are distinct in patients with cirrhosis compared with healthy individuals. By bacterial 16S rRNA gene sequencing, microbial diversity, and especially Bacteroidetes species, is shown to be



reduced in cirrhotic patients, while the number of species of Proteobacteria and Fusobacteria are increased [56]. In line with the previous study, *Bacteroides* has also been found to decrease at effective highly active anti-retroviral therapy (HAART), the diver- sity and composition of the fecal microbiota are not completely restored, and the dysbiosis remains [47]. South African teenage girls and young women have extremely high rates of HIV infection, a phenomenon that has been considered to be associated with biological factors. Recently, one study found that the vaginal microbiome might affect the risk of HIV infection. A bacterium in the vagina named *Prevotella bivia* has been identified as causing inflammation. A microbicidal gel is available that decomposes the anti-HIV drug tenofovir, thus leading to tenofovir treatment fail- ure [44]. Through close examination of the vaginal microbiome, Cohen [44] recently found an unusual bacterium named *Gard- nerella* in the vagina; this finding potentially explains the high infection rates in South African women and strongly suggests that the vaginal microbiome affects HIV risk. Cohen [44] found that *Gardnerella* "gobbles up" tenofovir, thus rapidly decreasing the levels of the drug and leading to tenofovir treatment failure.

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Acute-on-chronic liver failure (ACLF) syndrome is character- ized by the acute decompensation of cirrhosis, with high 28-d mortality. Based on the final clinical outcome at 90 d, we were the first to identify gut dysbiosis in ACLF patients, and to demon- strate its predictive value for mortality. We found a marked dif- ference between the gut microbiota of the ACLF group and that of the control group. Our study indicated that there are correlations between specific bacterial families and inflammatory cytokines in ACLF patients. We have demonstrated that the relative abun- dance of Pasteurellaceae and the model of end-stage liver disease (MELD) score are independent factors that predict the mortality rate, thus indicating that gut dysbiosis is associated with the mortality of patients with ACLF [59].

Although the exact reason for these changes in liver cirrho- sis remains unclear, these changes are certainly associated with reduced intestinal motility and pancreatobiliary secretions, an impaired intestinal barrier, and decreased gastric acidity. In ad- dition, 80% of hepatocellular carcinoma (HCC) develops in a mi- croenvironment of chronic injury, inflammation, or fibrosis [60]. Changes in the



composition of the gut microbiota promote HCC by contributing to hepatic inflammation through increased intes- tinal permeability and the activation of Toll-like receptors [60].

The incidence and prevalence of primary sclerosing cholangitis (PSC), PBC, and autoimmune hepatitis increases every year [61]. Autoimmune liver diseases are presumed to involve environmental factors in individuals with genetic susceptibility; however, the gut flora is relevant to pathogenesis. Recently, a study showed that patients with PSC-IBD have distinct gut microbiota and a significant increase in the abundance of *Escherichia*, Lachnos- piraceae, and *Megasphaera*, along with a near-absence of *Bacte- roides*, as compared with IBD patients and control patients [62]. Another study found patients with PBC-altered gut bacterial taxa that exhibited potential interactions through their associations with altered metabolism, immunity, and liver-function indicators [63]. There is evidence that bacterial antigens translocate across a leaky and inflamed gut wall into the portal and biliary system; thus, they may induce an abnormal immune response and initiate autoimmune liver disease [64].

NAFLD is a multifactorial disorder comprising a group of dis- eases. Genetic, epigenetic, and environmental factors interact with pose tissue inflammation, along with reduced insulin sensitivity, compared with mice that were fed with fish oil [110]. However, phenotypic differences between the dietary groups can be partly attributed to differences in microbiota composition. Increasing evidence shows that the gut microbiota is an important mod- ulator of the interaction between diet and the development of metabolic diseases [111]. Furthermore, recent studies have shown that the gut microbiota influences the circadian clock and under- goes circadian oscillations [112]. Disruption of the host circadian clock induces dysbiosis, which is associated with host metabolic disorders [113]. Obesity, which is associated with gut microbiota dysbiosis and altered metabolic pathways, induces impaired gut epithelial barrier function and has significant influences on phys- iological processes [114], such as gut and immune homeostasis [115], energy metabolism [116], acetate [25] and bile acid metab- olism [117], and intestinal hormone release [118].

Type 2 diabetes

T2D is a prevalent metabolic disease worldwide; the link be- tween the gut microbiome composition and the development of T2D is gradually being uncovered [119–121]. Growing numbers of studies indicate that an altered gut microbiome characterized by lower diversity and resilience is associated with diabetes. The mechanisms that cause the disease may be related to the trans- location of microbiota from the gut to the tissues, thus inducing inflammation [122]. Pedersen et al. [123] recently demonstrated that the human gut microbiome may affect the serum metab- olome and induce insulin resistance through species such as *Prevotella copri* and *Bacteroides vulgates*. Metformin is one of the most widely used antidiabetic drugs and is thought to confound the results of metagenomics data analysis [121]. The gut microbi- ota may directly affect T2D through its effect on the metabolism of amino acids; thus, future antidiabetic treatment strategies may target bacterial strains that cause imbalances in amino acid me- tabolism [121,124]. Therefore, obesity and its associated metabolic complications may be a result of complex gene-environment interactions. Microbiome interventions aimed at restoring the homeostasis of the gut microbiome have recently emerged, such as the ingestion of specific fibers or therapeutic microbes. These are promising strategies to reduce insulin resistance and related metabolic diseases.

The human microbiota and other diseases

Growing evidence indicates that alterations in the microbiota are implicated in the pathogenesis of a number of other dis- eases, such as severe asthma, food allergies, autism, and major depressive disorder (MDD) [125–130], all of which have recently received considerable scientific interest. Interestingly, these dis- eases may not involve direct interactions with the microbiota. However, the regulating function of the microbiota, such as the microbiota-gut-brain axis, may participate in the specific pathways of the diseases. The complex microbiota-host interactions are dynamic, involving a variety of mechanisms that include im-mune, hormonal, and neural pathways. Therefore, changes in the microbiota may result in the dysregulation of host homeostasis and in an increased susceptibility to these diseases. On the basis of these well-established connections between disease and the disruption of homeostatic interactions in the host, microbiota- targeted therapies may alter the community composition, and microbiota restoration might be used for treating these diseases.

The microbiota and allergic diseases

An early-life, antibiotic-driven low diversity in gut microbiota enhances susceptibility to allergic asthma [131], and thus may also affect asthma development in childhood after long-term follow-up. Of course, the mode, place of delivery, and infant feed- ing also affect the GI microbiota composition and subsequently influence the risk of atopic manifestations [132]. Bunyavanich et al. [128] found that infants with a gut microbiota enriched in Clostridia and Firmicutes at a host age of 3–6 months are



associ- ated with the resolution of cow's milk allergy (CMA) by the age of 8 years. Because the intestinal microbiota of an infant evolves rapidly in the first year, the early-life gut microbiota composition may be one of the determinants for CMA outcomes in childhood. The gut microbiota interacts with the immune system intimately, providing signals to promote the maturation of regulatory anti- gen-presenting cells and regulatory T cells (Tregs), which play a crucial role in the development of immunological tolerance. The specific members of the microbiota, such as *Clostridium* species, interact with Treg and regulate immunoglobulin E (IgE) levels [133]. Saarinen et al. [127] showed that the clinical course and prognosis of CMA are highly dependent on the milk-specific IgE status. A previous study also found that specific microbiotic sig- natures, such as that of *Clostridium sensu stricto*, can distinguish infants with IgE-mediated food allergies from those with non-IgE- mediated ones, and that *Clostridium sensu stricto* is positively cor- related with specific IgE level in serum [126].

The microbiota and psychiatric diseases

Psychiatric diseases have posed a severe threat to human health throughout history [134]. They are caused by a combination of bi-ological, psychological, and environmental factors [135-137]. The existence of a gut-brain axis has been acknowledged for decades. The gut-brain axis plays a key role in maintaining normal brain and GI function. More recently, the gut microbiota has emerged as a critical regulator of this axis. The concept of this axis has been extended to the "microbiota-gut-brain axis," and is now seen to involve a number of systems, including the endocrine sys- tem, neural system, metabolic system, and immune system, all of which are engaged in constant interaction [138]. Gut microbiota dysbiosis may increase the translocation of gut bacteria across the intestinal wall and into the mesenteric lymphoid tissue, thereby provoking an immune response that can lead to the release of inflammatory cytokines and the activation of the vagus nerve and spinal afferent neurons [139,140]. Autism spectrum disorder (ASD) has been reported as correlated with an altered gut microbiota, and low relative abundances of the mucolytic bacteria Akkerman- sia muciniphila and Bifidobacterium spp. have been found in the feces of children with autism [125]. Our previous study found an altered fecal microbiotic composition in patients with MDD. Most notably, the MDD groups had increased levels of Enterobacte- riaceae and Alistipes, but reduced levels of Faecalibacterium [130]. These studies suggest the role of the gut microbiota in autism and MDD as a part of the gut-brain axis; this suggested role should form a basis for further investigation of the combined effects of microbial, genetic, and hormonal changes in the development and clinical manifestation of autism and MDD.

Advancements in microbiota technology

Over the past few decades, human microbiome research has been revolutionized by high-throughput sequencing technology. High-throughput sequencing provides an opportunity for studies to focus on complex microbial systems without the need to clone individual genes. Initially, microbiota studies focused on compo- sitional studies (i.e., answering the question: what is there?) and functional studies (i.e., what are they doing?). With the develop- ment of sequencing technology and bioinformatics analysis, it has become increasingly interesting to study the activity of microbes within microbial communities. It is widely accepted that the mi- crobes with the highest abundance are not always the most active ones. RNA sequencing (RNAseq) permits the analysis of gene ex- pression, adding valuable expression data to compositional data sets. Gosalbes et al. [141] performed the first metatranscriptomic analysis of the healthy human gut microbiota in 2011. The anal- ysis of 16S transcripts showed the phylogenetic structure of the active microbial community. Lachnospiraceae, Ruminococcaceae, Bacteroidaceae, Prevotellaceae, and Rickenellaceae were the pre-dominant families detected in the active microbiota. The primary functional roles of the gut microbiota were found to be carbohy- drate metabolism, energy production, and the synthesis of cellu- lar components. A systematic comparison of the gut metagenome and metatranscriptome revealed that a substantial fraction (41%) of microbial transcripts was not differentially regulated relative to their genomic abundances. The metatranscriptional profiles were significantly more individualized than the DNA-level functional profiles but were less variable in their microbial composition [142]. A transcriptome analysis of bacteriophage communities in the periodontal microbiota was recently performed using RNAseq. Oral phages were found to be more highly expressed in individuals with relative periodontal health [143].

To achieve precise microbiome-based medicine in the future, it is necessary to understand which individual microorganisms mediate vital microbiome-host interaction(s) under health or disease conditions. Most gut microbes are currently uncultivable. Even with the use of recent technologies, such as gnotobiotic mice and anaerobic culturing techniques, it is possible to culture only approximately half of the bacterial species identified by 16S rDNA high-throughput sequencing [144]. In addition, species- level identification may not reflect the real situation because most of the functional diversity can be reflected only at the strain level. Therefore, it will be crucial to develop technologies to iden- tify and isolate these microorganisms and/or microbial consortia.

Compared with traditional microbiology approaches, the use of anaerobic conditions and gnotobiotic animals largely facilitates the cultivation of difficult-to-grow microbes. Numerous



previously uncultivable microbes can now be cultured in a laboratory set- ting [145]. A chip-based isolation device (the iChip) was recently developed and was specifically designed to identify uncultivable microbes within complex microbial ecosystems [146]. The iChip is composed of hundreds of miniature diffusion chambers, each of which is inoculated with a single environmental cell. The ca- pacity for microbial recovery using the iChip is many times high- er than that of standard cultivation, and the resulting species are of significant phylogenetic novelty [146]. A new device for *in situ* cultivation (the I-tip) was subsequently developed. The principal of the I-tip is similar to that of the iChip; however, the I-tip traps individual microbes within a gel, thus allowing for the passage of metabolites and nutrients. The *in situ* isolation of microbes from invertebrate organisms using the I-tip has recovered isolates from 34 novel microbial species [147].

Simulating GIT conditions can greatly facilitate *in vitro* cultiva- tion. The Simulator of the Human Intestinal Microbial Ecosystem (SHIME) has succeeded in establishing stable, reactor-grown GIT microbial communities. More importantly, this system is able to precisely simulate different regions of the human GIT, thus allowing the diversity of the community to be studied *in vitro* in a different niche. The power of SHIME has been estimated by numerous studies [148–150]. For example, it has been found that different regions of SHIME are colonized by different unique microbial communities when cultured with microbes. The distri- bution is highly similar to that of the living host, such as the prev- alence of *Bacterioides/Prevotella* spp. and *Lactobacillus* spp. in the colon [148]. Identifying single cells that produce metabolites of interest within complex microbial ecosystems is very important for un- derstanding microbiome-host interactions. With this purpose, a flexible high-throughput approach using a combination of micro- fluidics and fluorescence-activated cell sorting has recently been developed [151]. This system has successfully identified xylose- overconsuming *Saccharomyces cerevisiae* and *L*-lactate-producing

E. coli cells from a population. This system also allows for the screening of mutants in known pathways.

The use of traditional animal models in microbiome studies continues to provide insight into hostmicrobiota interactions. However, animal models often do not predict the results obtained in humans, thus posing a particular problem when considering challenges relating to the oral absorption of drugs and nutrients. Here, we introduce two methods of in vitro simulation based on stem cells: the gut-on-achip system and colonic stem cell con- struction. The gut-on-a-chip system takes advantage of biomaterial engineering and provides an optional approach to study the com- plex interactions occurring within the gut microbiome. This system is an *in vitro* living cell-based model of the intestine that mimics the properties of the human gut along with crucial microbial sym- bionts. Biomimetic human gut-on-achip micro-devices are usually composed of microfluidic channels and a porous flexible mem- brane that are coated with an extracellular matrix and lined with human intestinal epithelial cells [152]; such devices mimic the complex structure and physiology of a living intestine. Microfluidic devices can also be used to study microbe-microbe interactions, such as chemotaxis/chemical attraction and quorum sensing [153]; such interactions have been shown to be more effectively studied using microfluidic devices than using traditional capillary-based assays [154]. In addition, given the recapitulation of many complex functions of the normal human intestine, it may also become an essential platform for drug screening and toxicology testing.

Colonic stem cell construction is a recently developed *in vitro* system that is used to grow 3D organoid colonic epithelium structures that are guided by microstructures without the utilization of microfluidics technology [155]. Within a Matrigel overlay, spherical 3D structures grown from colonic stem cells or intesti- nal stem cells are collected from an array containing individually grown structures. These membrane-free 3D stem-cell-derived organoids, which contain various differentiated cell types, form a barrier similar to that of intestinal or colonic epithelia [156,157]. These organoids have recently been used to demonstrate that the *Salmonella enterica* serovar Typhimurium can successfully invade the epithelial cell layer [157], and that *C. difficile* can disrupt the epithelial barrier function [158]. This technology also provides more novel and valuable methods for higher throughput micro- biome studies than existing models, although this technique is still in its infancy.

APPLICATION OF THE HUMAN MICROBIOTA

The human microbiome can be considered as an important origin of resources for genetic diversity, a modifier of disease, an essential component of immunity, and a functional entity that influences metabolism and modulates drug interactions. On one hand, there are many potential probiotics or beneficial bacteria that may prevent or treat certain diseases, although most of them cannot be cultivated at present [159]. For example, some of these gut microbes belong to genera that contain many probiotics such as *Lactobacillus* and *Bifidobacterium*. Some are novel potentially beneficial



bacteria, such as *Faecalibacterium prausnitzii* for treat- ing IBD and irritable bowel syndrome (IBS), and *Akkermansia muciniphila* for improving metabolic health [160]. On the other hand, as our second genome, the human microbiome must pro- duce a large number of metabolites. Some isolated metabolites have important potential applications, although it still remains a great challenge to isolate and identify all the metabolites of the human microbiome. For example, Chu et al. [161] discovered methicillin-resistant *Staphylococcus aureus*-active antibiotics by using primary sequencing from the human microbiome.

With the increased understanding of the relationship between the human microbiome and a variety of diseases, the use of these findings to predict or diagnose diseases has attracted a great deal of attention [162]. Enrichments of some microbes are noted as potential biomarkers in some research; however, these alterations are often observed in other research as well, and cannot be distinguished among different diseases. In contrast, clinical models based on tens of genes within a metagenome analysis perform better in diagnostics and predicting diseases. In addition, we found that the *Bifidobacterium*/Enterobacteriaceae (B/E) ratio indicates the microbial colonization resistance of the bowel, and that this ratio is considered to be an indicator of human microbiome heath. The B/E ratio is higher than 1 in people with healthy microbiomes, whereas it is far below 1 in patients with cirrhosis and patients with the avian influenza H7N9 infections [163,164].

The prevention and treatment of diseases by targeting the microbiome have been widely investigated, and some therapies have been successfully applied in the clinic. The administration of probiotics is reported to help restore the health of H7N9 patients more quickly [164]. Fecal microbiome transplantation has exhib- ited better clinical efficacy than antibiotics in the treatment of C. difficile infections [165]. Substantial progress has also been made in the treatment of liver diseases by modulating the gut micro- biome. A clinical trial showed that probiotic VSL#3 reduces liver disease severity and hospitalization in patients with cirrhosis [166]; the administration of Lactobacillus salivarius LI01 or Pedi- ococcus pentosaceus LI05 improves the acute liver injury induced by Dgalactosamine in rats [167]. Furthermore, the regulation of the human microbiome plays important roles in the treatment of GI diseases, such as infectious diarrhea, antibiotic-associated diarrhea, inflammatory bowel disease, and necrotizing enter- ocolitis. For example, the oral administration of a mixture of 17 Clostridia strains from the human microbiota to adult mice was found to attenuate disease in models of colitis and allergic diarrhea [168]. Modulation of the gut microbiome may also con- tribute to the treatment of cancer. Iida et al. [169] reported that optimal responses to cancer therapy require an intact commensal microbiota that mediates the therapy effects by modulating mye-loidderived cell functions in the tumor microenvironment. Viaud et al. [170] reported that the gut microbiota helps to shape the anticancer immune response of cyclophosphamide. In addition, many clinical studies have shown that probiotics and their prod- ucts have outstanding effects on the treatment of allergic diseas- es, especially those in infants [171].

FUTURE PERSPECTIVES

The human microbiota plays an important role in the well-being of the human host, and participates actively in the develop- ment of a wide variety of diseases. Given the extensive influence of microorganisms throughout the human body, we propose that research on host-microbiota interactions should go beyond a char- acterization of the community composition and an investigation of the community members' associations. From the structure to the function of the microbiota, future research should move mi- crobiome investigations toward providing explanations of causal- ity. With new techniques for microbiota function prediction, new microbiota interaction models, and novel analytical and simulation approaches, future advances will help to clarify the interactions between the microbiota and human development, and the po- tential roles of those microbiota involved in the mechanisms of various diseases, such as liver diseases, bacterial infection, cancer, psychiatric diseases, and metabolic diseases. The crucial roles of the human microbiota should be investigated at a much deeper level, and microbiome-based diagnosis and treatment strategies will be used for future personalized medicine work.

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COMPLIANCE WITH ETHICS GUIDELINES

Baohong Wang, Mingfei Yao, Longxian Lv, Zongxin Ling, and Lanjuan Li declare that they have no conflict of interest or finan- cial conflicts to disclose.

REFERENCES

- [1]. O'Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Rep 2006;7(7):688–93.
- [2]. Ursell LK, Haiser HJ, Van Treuren W, Garg N, Reddivari L, Vanamala J, et al. The intestinal metabolome: an intersection between microbiota and host. Gastroenterology 2014;146(6):1470–6.
- [3]. Whitman WB, Coleman DC, Wiebe WJ. Prokaryotes: the unseen majority. Proc Natl Acad Sci USA 1998:95(12):6578–83.
- [4]. Savage DC. Microbial ecology of the gastrointestinal tract. Annu Rev Microbiol 1977;31(1):107–33.
- [5]. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell 2006;124(4):837–48.
- [6]. Wang B, Li L. Who determines the outcomes of HBV exposure? Trends Mi-crobiol 2015;23(6):328–29.
- [7]. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut mi- crobes associated with obesity. Nature 2006;444(7122):1022–3.
- [8]. Wang B, Jiang X, Cao M, Ge J, Bao Q, Tang L, et al. Altered fecal microbiota correlates with liver biochemistry in nonobese patients with non-alcoholic fatty liver disease. Sci Rep 2016;6:32002.
- [9]. Gill SR, Pop M, Deboy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, et al. Metagenomic analysis of the human distal gut microbiome. Science 2006;312(5778):1355–9.
- [10]. Roberfroid MB, Bornet F, Bouley C, Cummings JH. Colonic microflora: nutri- tion and health. Summary and conclusions of an International Life Scienc- es Institute (ILSI) [Europe] workshop held in Barcelona, Spain. Nutr Rev 1995;53(5):127–30.
- [11]. Cash HL, Whitham CV, Behrendt CL, Hooper LV. Symbiotic bacteria direct expression of an intestinal bactericidal lectin. Science 2006;313(5790):1126–30.
- [12]. Hooper LV, Stappenbeck TS, Hong CV, Gordon JI. Angiogenins: a new class of microbicidal proteins involved in innate immunity. Nat Immunol 2003;4(3):269–73.
- [13]. Schauber J, Svanholm C, Termén S, Iffland K, Menzel T, Scheppach W, et al. Expression of the cathelicidin LL-37 is modulated by short chain fatty acids in colonocytes: relevance of signalling pathways. Gut 2003;52(5):735–41.
- [14]. Bouskra D, Brézillon C, Bérard M, Werts C, Varona R, Boneca IG, et al. Lym- phoid tissue genesis induced by commensals through NOD1 regulates intes- tinal homeostasis. Nature 2008;456(7221):507–10.
- [15]. Rakoff-Nahoum S, Medzhitov R. Innate immune recognition of the indige- nous microbial flora. Mucosal Immunol 2008;1(Suppl 1):S10–4.
- [16]. Macpherson AJ, Harris NL. Interactions between commensal intestinal bac- teria and the immune system. Nat Rev Immunol 2004;4(6):478–85.
- [17]. Sekirov I, Russell SL, Antunes LC, Finlay BB. Gut microbiota in health and dis-ease. Physiol Rev 2010;90(3):859–904.
- [18]. Sartor RB. Microbial influences in inflammatory bowel diseases. Gastroen-terology 2008;134(2):577–94.
- [19]. Liu Q, Duan Z, Ha D, Bengmark S, Kurtovic J, Riordan SM. Synbiotic modula- tion of gut flora: effect on minimal hepatic encephalopathy in patients with cirrhosis. Hepatology 2004;39(5):1441–9. Scanlan PD, Shanahan F, Clune Y, Collins JK, O'Sullivan GC, O'Riordan M, et al. Culture-independent analysis of the gut microbiota in colorectal cancer and polyposis. Environ Microbiol 2008;10(3):789–98.
- [20]. Verhulst SL, Vael C, Beunckens C, Nelen V, Goossens H, Desager K. A longitu-dinal analysis on the association between antibiotic use, intestinal microflo- ra, and wheezing during the first year of life. J Asthma 2008;45(9):828–32.
- [21]. Finegold SM, Molitoris D, Song Y, Liu C, Vaisanen ML, Bolte E, et al. Gastrointestinal microflora studies in late-onset autism. Clin Infect Dis 2002;35(Sup- pl 1):S6–16.
- [22]. Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, et al. Innate immunity and intestinal microbiota in the development of Type 1 diabetes. Nature 2008;455(7216):1109–13.
- [23]. Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy har- vest. Nature 2006;444(7122):1027–31.
- [24]. Perry RJ, Peng L, Barry NA, Cline GW, Zhang D, Cardone RL, et al. Acetate mediates a



- microbiome-brain-β-cell axis to promote metabolic syndrome. Nature 2016;534(7606):213–7.
- [25]. Cani PD, Dewever C, Delzenne NM. Inulin-type fructans modulate gastroin- testinal peptides involved in appetite regulation (glucagon-like peptide-1 and ghrelin) in rats. Br J Nutr 2004;92(3):521-6.
- [26]. Hollister EB, Gao C, Versalovic J. Compositional and functional features of the gastrointestinal microbiome and their effects on human health. Gastro- enterology 2014;146(6):1449–58.
- [27]. Rogier EW, Frantz AL, Bruno ME, Wedlund L, Cohen DA, Stromberg AJ, et al. Lessons from mother: long-term impact of antibodies in breast milk on the gut microbiota and intestinal immune system of breastfed offspring. Gut Microbes 2014;5(5):663–8.
- [28]. Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune re-sponses during health and disease. Nat Rev Immunol 2009;9(5):313–23.
- [29]. Larsbrink J, Rogers TE, Hemsworth GR, McKee LS, Tauzin AS, Spadiut O, et al. A discrete genetic locus confers xyloglucan metabolism in select human gut Bacteroidetes. Nature 2014;506(7489):498–502.
- [30]. Goh YJ, Klaenhammer TR. Genetic mechanisms of prebiotic oligosaccharide metabolism in probiotic microbes. Annu Rev Food Sci Technol 2015;6:137–56.
- [31]. Morowitz MJ, Carlisle EM, Alverdy JC. Contributions of intestinal bacte- ria to nutrition and metabolism in the critically ill. Surg Clin North Am 2011;91(4):771–85.
- [32]. Duncan SH, Louis P, Thomson JM, Flint HJ. The role of pH in determining the species composition of the human colonic microbiota. Environ Microbiol 2009;11(8):2112–22.
- [33]. Cani PD, Everard A, Duparc T. Gut microbiota, enteroendocrine functions and metabolism. Curr Opin Pharmacol 2013;13(6):935–40.
- [34]. Flint HJ, Scott KP, Louis P, Duncan SH. The role of the gut microbiota in nutri- tion and health. Nat Rev Gastroenterol Hepatol 2012;9(10):577–89.
- [35]. Kang Z, Zhang J, Zhou J, Qi Q, Du G, Chen J. Recent advances in microbi- al production of δ-aminolevulinic acid and vitamin B₁₂. Biotechnol Adv 2012;30(6):1533–42.
- [36]. Cebra JJ. Influences of microbiota on intestinal immune system develop- ment. Am J Clin Nutr 1999;69(5):1046S-51S.
- [37]. Thaiss CA, Zmora N, Levy M, Elinav E. The microbiome and innate immunity. Nature 2016;535(7610):65–74.
- [38]. Madsen KL, Doyle JS, Jewell LD, Tavernini MM, Fedorak RN. *Lactobacillus* spe- cies prevents colitis in interleukin 10 gene-deficient mice. Gastroenterology 1999;116(5):1107–14.
- [39]. Magrone T, Jirillo E. The interplay between the gut immune system and microbiota in health and disease: nutraceutical intervention for restoring intestinal homeostasis. Curr Pharm Des 2013;19(7):1329–42.
- [40]. Brenchley JM, Douek DC. Microbial translocation across the GI tract. Annu Rev Immunol 2012;30:149–73.
- [41]. Hand TW, Dos Santos LM, Bouladoux N, Molloy MJ, Pagán AJ, Pepper M, et al. Acute gastrointestinal infection induces long-lived microbiota-specific T cell responses. Science 2012;337(6101):1553–6.
- [42]. Ling Z, Liu X, Cheng Y, Jiang X, Jiang H, Wang Y, et al. Decreased diversity of the oral microbiota of patients with hepatitis B virus-induced chronic liver disease: a pilot report. Sci Rep 2015;5:17098.
- [43]. Cohen J. Vaginal microbiome affects HIV risk. Science 2016;353(6297):331.
- [44]. Gu S, Chen Y, Zhang X, Lu H, Lv T, Shen P, et al. Identification of key taxa that favor intestinal colonization of *Clostridium difficile* in an adult Chinese population. Microbes Infect 2016;18(1):30–8.
- [45]. Ling Z, Liu X, Jia X, Cheng Y, Luo Y, Yuan L, et al. Impacts of infection with different toxigenic *Clostridium difficile* strains on faecal microbiota in children. Sci Rep 2014;4:7485.
- [46]. Ling Z, Jin C, Xie T, Cheng Y, Li L, Wu N. Alterations in the fecal microbiota of patients with HIV-1 infection: an observational study in a Chinese popula- tion. Sci Rep 2016;6:30673.
- [47]. Xu M, Wang B, Fu Y, Chen Y, Yang F, Lu H, et al. Changes of fecal *Bifidobacterium* species in adult patients with hepatitis B virus-induced chronic liver disease. Microb Ecol 2012;63(2):304–13
- [48]. Hu Z, Zhang Y, Li Z, Yu Y, Kang W, Han Y, et al. Effect of *Helicobacter pylori* in-fection on chronic periodontitis by the change of microecology and inflam-mation. Oncotarget 2016;7(41):66700–12.
- [49]. Ling Z, Kong J, Liu F, Zhu H, Chen X, Wang Y, et al. Molecular analysis of the diversity of vaginal microbiota associated with bacterial vaginosis. BMC
- [50]. Genomics 2010;11:488.
- [51]. Giannelli V, Di Gregorio V, Iebba V, Giusto M, Schippa S, Merli M, et al. Mi- crobiota and the gut-liver axis: bacterial translocation, inflammation and infection in cirrhosis. World J Gastroenterol 2014;20(45):16795–810.



- [52]. Nardone G, Rocco A. Probiotics: a potential target for the prevention and treatment of steatohepatitis. J Clin Gastroenterol 2004;38(Suppl 2):S121–2.
- [53]. Cesaro C, Tiso A, Del Prete A, Cariello R, Tuccillo C, Cotticelli G, et al. Gut microbiota and probiotics in chronic liver diseases. Dig Liver Dis 2011;43(6): 431–8.
- [54]. Lakshmi CP, Ghoshal UC, Kumar S, Goel A, Misra A, Mohindra S, et al. Fre- quency and factors associated with small intestinal bacterial overgrowth in patients with cirrhosis of the liver and extra hepatic portal venous obstruction. Dig Dis Sci 2010;55(4):1142–8.
- [55]. Gupta A, Dhiman RK, Kumari S, Rana S, Agarwal R, Duseja A, et al. Role of small intestinal bacterial overgrowth and delayed gastrointestinal transit time in cirrhotic patients with minimal hepatic encephalopathy. J Hepatol 2010;53(5):849–55.
- [56]. Chen Y, Yang F, Lu H, Wang B, Chen Y, Lei D, et al. Characterization of fe- cal microbial communities in patients with liver cirrhosis. Hepatology 2011;54(2):562–72.
- [57]. Qin N, Yang F, Li A, Prifti E, Chen Y, Shao L, et al. Alterations of the human gut microbiome in liver cirrhosis. Nature 2014;513(7516):59–64.
- [58]. Chen Y, Ji F, Guo J, Shi D, Fang D, Li L. Dysbiosis of small intestinal microbiota in liver cirrhosis and its association with etiology. Sci Rep 2016;6:34055.
- [59]. Chen Y, Guo J, Qian G, Fang D, Shi D, Guo L, et al. Gut dysbiosis in acute-on- chronic liver failure and its predictive value for mortality. J Gastroenterol Hepatol 2015;30(9):1429–37.
- [60]. Dapito DH, Mencin A, Gwak GY, Pradere JP, Jang MK, Mederacke I, et al. Pro- motion of hepatocellular carcinoma by the intestinal microbiota and TLR4. Cancer Cell 2012;21(4):504–16.
- [61]. Ozaslan E, Efe C. Further considerations in autoimmune hepatitis. J Hepatol 2016;64(6):1457–8.
- [62]. Marchesi JR, Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, et al. The gut microbiota and host health: a new clinical frontier. Gut 2016;65(2): 330–9.
- [63]. Lv L, Fang D, Shi D, Chen D, Yan R, Zhu Y, et al. Alterations and correlations of the gut microbiome, metabolism and immunity in patients with primary biliary cirrhosis. Environ Microbiol 2016;18(7):2272–86.
- [64]. Björnsson E, Cederborg A, Åkvist A, Simren M, Stotzer PO, Bjarnason I. Intestinal permeability and bacterial growth of the small bowel in patients with primary sclerosing cholangitis. Scand J Gastroenterol 2005;40(9):1090–4.
- [65]. Tilg H, Moschen AR, Roden M. NAFLD and diabetes mellitus. Nat Rev Gastro- enterol Hepatol 2017;14(1):32–42.
- [66]. Wesolowski SR, Kasmi KC, Jonscher KR, Friedman JE. Developmental origins of NAFLD: a womb with a clue. Nat Rev Gastroenterol Hepatol. Epub 2016 Oct 26.
- [67]. Boursier J, Mueller O, Barret M, Machado M, Fizanne L, Araujo-Perez F, et al. The severity of nonalcoholic fatty liver disease is associated with gut dys- biosis and shift in the metabolic function of the gut microbiota. Hepatology 2016;63(3):764–75.
- [68]. de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, et al. Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. Lancet Oncol 2012;13(6):607– 15.
- [69]. Wong BCY, Lam SK, Wong WM, Chen JS, Zheng TT, Feng RE, et al.; China Gas- tric Cancer Study Group. *Helicobacter pylori* eradication to prevent gastric cancer in a high-risk region of China: a randomized controlled trial. JAMA 2004;291(2):187–94.
- [70]. Herrera V, Parsonnet J. *Helicobacter pylori* and gastric adenocarcinoma. Clin Microbiol Infect 2009;15(11):971–6.
- [71]. El-Omar EM, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, et al. Interleukin-1 polymorphisms associated with increased risk of gastric can- cer. Nature 2000;404(6776):398–402.
- [72]. de Sablet T, Piazuelo MB, Shaffer CL, Schneider BG, Asim M, Chaturvedi R, et al. Phylogeographic origin of *Helicobacter pylori* is a determinant of gastric cancer risk. Gut 2011;60(9):1189–95.
- [73]. Rhead JL, Letley DP, Mohammadi M, Hussein N, Mohagheghi MA, Eshagh Hosseini M, et al. A new *Helicobacter pylori* vacuolating cytotoxin determinant, the intermediate region, is associated with gastric cancer. Gastroenter-ology 2007;133(3):926–36.
- [74]. Cover TL, Krishna US, Israel DA, Peek RM Jr. Induction of gastric epitheli- al cell apoptosis by *Helicobacter pylori* vacuolating cytotoxin. Cancer Res 2003;63(5):951–7.
- [75]. Oertli M, Sundquist M, Hitzler I, Engler DB, Arnold IC, Reuter S, et al. DC- derived IL-18 drives Treg differentiation, murine *Helicobacter pylori*-specific immune tolerance, and asthma protection. J Clin Invest 2012;122(3):1082–96.
- [76]. Blaser MJ, Perez-Perez GI, Kleanthous H, Cover TL, Peek RM, Chyou PH, et al. Infection with *Helicobacter pylori* strains possessing *cagA* is associated with an increased risk of developing adenocarcinoma of the stomach. Cancer Res 1995;55(10):2111–5.
- [77]. Kaparakis M, Turnbull L, Carneiro L, Firth S, Coleman HA, Parkington HC, et al. Bacterial membrane vesicles deliver peptidoglycan to NOD1 in epithelial cells. Cell Microbiol



- 2010;12(3):372-85.
- [78]. Odenbreit S, Püls J, Sedlmaier B, Gerland E, Fischer W, Haas R. Translocation of *Helicobacter pylori cagA* into gastric epithelial cells by type IV secretion. Science 2000;287(5457):1497–500.
- [79]. Odenbreit S, Püls J, Sedlmaier B, Gerland E, Fischer W, Haas R. Translocation of *Helicobacter pylori cagA* into gastric epithelial cells by type IV secretion. Science 2000;287(5457):1497–500.
- [80]. Lertpiriyapong K, Whary MT, Muthupalani S, Lofgren JL, Gamazon ER, Feng Y, et al. Gastric colonisation with a restricted commensal microbiota repli- cates the promotion of neoplastic lesions by diverse intestinal microbiota in the Helicobacter pylori INS-GAS mouse model of gastric carcinogenesis. Gut 2014;63(1):54-63.
- [81]. Sanapareddy N, Legge RM, Jovov B, McCoy A, Burcal L, Araujo-Perez F, et al. Increased rectal microbial richness is associated with the presence of colorectal adenomas in humans. ISME J 2012;6(10):1858–68.
- [82]. Castellarin M, Warren RL, Freeman JD, Dreolini L, Krzywinski M, Strauss J, et al. *Fusobacterium nucleatum* infection is prevalent in human colorectal carci- noma. Genome Res 2012;22(2):299–306.
- [83]. Zackular JP, Baxter NT, Iverson KD, Sadler WD, Petrosino JF, Chen GY, et al. The gut microbiome modulates colon tumorigenesis. MBio 2013;4(6): e00692–13.
- [84]. Keku TO, McCoy AN, Azcarate-Peril AM. *Fusobacterium* spp. and colorectal cancer: cause or consequence? Trends Microbiol 2013;21(10):506–8.
- [85]. Feng Q, Liang S, Jia H, Stadlmayr A, Tang L, Lan Z, et al. Gut microbiome de- velopment along the colorectal adenoma-carcinoma sequence. Nat Commun 2015;6:6528.
- [86]. Rubinstein MR, Wang X, Liu W, Hao Y, Cai G, Han YW. *Fusobacterium nuclea- tum* promotes colorectal carcinogenesis by modulating E-cadherin/β-catenin signaling via its FadA adhesin. Cell Host Microbe 2013;14(2):195–206.
- [87]. Kostic AD, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, et al. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. Cell Host Microbe 2013;14(2): 207–15.
- [88]. Sokol SY. Wnt signaling and dorso-ventral axis specification in vertebrates. Curr Opin Genet Dev 1999;9(4):405–10.
- [89]. Sears CL. Enterotoxigenic *Bacteroides fragilis*: a rogue among symbiotes. Clin Microbiol Rev 2009;22(2):349–69.
- [90]. Shiryaev SA, Remacle AG, Chernov AV, Golubkov VS, Motamedchaboki K, Muranaka N, et al. Substrate cleavage profiling suggests a distinct function of *Bacteroides fragilis* metalloproteinases (fragilysin and metalloproteinase II) at the microbiome-inflammation-cancer interface. J Biol Chem 2013;288(48): 34956–67.
- [91]. Wu S, Rhee KJ, Albesiano E, Rabizadeh S, Wu X, Yen HR, et al. A human co-lonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. Nat Med 2009;15(9):1016–22.
- [92]. Huycke MM, Abrams V, Moore DR. *Enterococcus faecalis* produces extracel- lular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. Carcinogenesis 2002;23(3):529–36.
- [93]. Cuevas-Ramos G, Petit CR, Marcq I, Boury M, Oswald E, Nougayrède JP. *Es- cherichia coli* induces DNA damage *in vivo* and triggers genomic instability in mammalian cells. Proc Natl Acad Sci USA 2010;107(25):11537–42.
- [94]. Howe GR, Benito E, Castelleto R, Cornée J, Estève J, Gallagher RP, et al. Die- tary intake of fiber and decreased risk of cancers of the colon and rectum: evidence from the combined analysis of 13 case-control studies. J Natl Can- cer Inst 1992;84(24):1887–96.
- [95]. Clausen MR, Bonnén H, Mortensen PB. Colonic fermentation of dietary fibre to short chain fatty acids in patients with adenomatous polyps and colonic cancer. Gut 1991;32(8):923–8.
- [96]. Singh N, Gurav A, Sivaprakasam S, Brady E, Padia R, Shi H, et al. Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. Immunity 2014; 40(1):128–39.
- [97]. Lagergren J, Bergström R, Lindgren A, Nyrén O. Symptomatic gastroesoph- ageal reflux as a risk factor for esophageal adenocarcinoma. N Engl J Med 1999;340(11):825–31.
- [98]. Lagergren J. Adenocarcinoma of oesophagus: what exactly is the size of the problem and who is at risk? Gut 2005;54(Suppl 1):i1–5.
- [99]. Anderson LA, Murphy SJ, Johnston BT, Watson RG, Ferguson HR, Bamford KB, et al. Relationship between *Helicobacter pylori* infection and gastric atrophy and the stages of the oesophageal inflammation, metaplasia, ad-enocarcinoma sequence: results from the FINBAR case-control study. Gut 2008;57(6):734–9.
- [100].Pei Z, Bini EJ, Yang L, Zhou M, Francois F, Blaser MJ. Bacterial biota in the hu-man distal esophagus. Proc Natl Acad Sci USA 2004;101(12):4250–5.



- [101]. Yang L, Lu X, Nossa CW, Francois F, Peek RM, Pei Z. Inflammation and intesti- nal metaplasia of the distal esophagus are associated with alterations in the microbiome. Gastroenterology 2009;137(2):588–97.
- [102]. Finlay IG, Wright PA, Menzies T, McArdle CS. Microbial flora in carcinoma of oesophagus. Thorax 1982;37(3):181–4.
- [103].El-Serag HB, Sonnenberg A. Opposing time trends of peptic ulcer and reflux disease. Gut 1998;43(3):327–33.
- [104].Hamada H, Haruma K, Mihara M, Kamada T, Yoshihara M, Sumii K, et al. High incidence of reflux oesophagitis after eradication therapy for *Helicobacter pylori*: impacts of hiatal hernia and corpus gastritis. Aliment Pharmacol Ther 2000;14(6):729–35.
- [105].Sommer F, Bäckhed F. The gut microbiota—masters of host development and physiology. Nat Rev Microbiol 2013;11(4):227–38.
- [106].Franks PW, McCarthy MI. Exposing the exposures responsible for type 2 dia- betes and obesity. Science 2016;354(6308):69–73.
- [107].Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, et al.; MetaHIT Consortium. Richness of human gut microbiome correlates with metabolic markers. Nature 2013;500(7464):541–6
- [108]. Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. Nature 2009; 457(7228): 480–4.
- [109].Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. Science 2013;341(6150):1241214.
- [110].David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, et al. Diet rapidly and reproducibly alters the human gut microbiome. Nature 2014;505(7484):559–63.
- [111].Caesar R, Tremaroli V, Kovatcheva-Datchary P, Cani PD, Bäckhed F. Crosstalk between gut microbiota and dietary lipids aggravates WAT inflammation through TLR signaling. Cell Metab 2015;22(4):658–68.
- [112]. Sonnenburg JL, Bäckhed F. Diet-microbiota interactions as moderators of human metabolism. Nature 2016;535(7610):56–64.
- [113].Leone V, Gibbons SM, Martinez K, Hutchison AL, Huang EY, Cham CM, et al. Effects of diurnal variation of gut microbes and high-fat feeding on host cir- cadian clock function and metabolism. Cell Host Microbe 2015;17(5):681–9.
- [114]. Thaiss CA, Zeevi D, Levy M, Zilberman-Schapira G, Suez J, Tengeler AC, et al. Transkingdom control of microbiota diurnal oscillations promotes metabolic homeostasis. Cell 2014;159(3):514– 29
- [115].Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. Nature 2012;489(7415):242–9.de Vos WM, Nieuwdorp M. Genomics: a gut prediction. Nature 2013;498(7452): 48–9.
- [116].Delzenne NM, Cani PD. Gut microflora is a key player in host energy homeo-stasis. Med Sci (Paris) 2008;24(5):505–10.
- [117]. Wahlström A, Sayin SI, Marschall HU, Bäckhed F. Intestinal crosstalk between bile acids and microbiota and its impact on host metabolism. Cell Metab 2016;24(1):41–50.
- [118].Camilleri M. Peripheral mechanisms in appetite regulation. Gastroenterology 2015;148(6):1219–33.
- [119].Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. Nature 2012;490(7418):55–60.
- [120].Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. Nature 2013;498(7452):99–103.
- [121].Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, et al.; MetaHIT Consortium. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. Nature 2015;528(7581): 262–6.
- [122].Burcelin R. Gut microbiota and immune crosstalk in metabolic disease. Mol Metab 2016;5(9):771–81.
- [123].Pedersen HK, Gudmundsdottir V, Nielsen HB, Hyotylainen T, Nielsen T, Jensen BA, et al.; MetaHIT Consortium. Human gut microbes impact host serum metabolome and insulin sensitivity. Nature 2016;535(7612):376–81.
- [124].Mardinoglu A, Boren J, Smith U. Confounding effects of metformin on the human gut microbiome in type 2 diabetes. Cell Metab 2016;23(1):10–2.
- [125]. Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA. Low relative abundances of the mucolytic bacterium *Akkermansia muciniphila* and *Bifidobacterium* spp. in feces of children with autism. Appl Environ Mi- crobiol 2011;77(18):6718–21.
- [126].Ling Z, Li Z, Liu X, Cheng Y, Luo Y, Tong X, et al. Altered fecal microbiota composition



- associated with food allergy in infants. Appl Environ Microbiol 2014;80(8):2546-54.
- [127].Saarinen KM, Pelkonen AS, Mäkelä MJ, Savilahti E. Clinical course and prog-nosis of cow's milk allergy are dependent on milk-specific IgE status. J Aller- gy Clin Immunol 2005;116(4):869–75.
- [128].Bunyavanich S, Shen N, Grishin A, Wood R, Burks W, Dawson P, et al. Early- life gut microbiome composition and milk allergy resolution. J Allergy Clin Immunol 2016;138(4):1122–30.
- [129].Tomova A, Husarova V, Lakatosova S, Bakos J, Vlkova B, Babinska K, et al. Gastrointestinal microbiota in children with autism in Slovakia. Physiol Behav 2015;138:179–87.
- [130]. Jiang H, Ling Z, Zhang Y, Mao H, Ma Z, Yin Y, et al. Altered fecal microbiota composition in patients with major depressive disorder. Brain Behav Immun 2015;48:186–94.
- [131].Russell SL, Gold MJ, Hartmann M, Willing BP, Thorson L, Wlodarska M, et al. Early life antibiotic-driven changes in microbiota enhance susceptibility to allergic asthma. EMBO Rep 2012;13(5):440–7.
- [132].van Nimwegen FA, Penders J, Stobberingh EE, Postma DS, Koppelman GH, Kerkhof M, et al. Mode and place of delivery, gastrointestinal micro- biota, and their influence on asthma and atopy. J Allergy Clin Immunol 2011;128(5):948–55.e1–3.
- [133]. Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, et al. Induction of colonic regulatory T cells by indigenous *Clostridium* species. Science 2011;331(6015):337–41.
- [134].Charlson FJ, Baxter AJ, Cheng HG, Shidhaye R, Whiteford HA. The burden of mental, neurological, and substance use disorders in China and India: a systematic analysis of community representative epidemiological studies. Lancet 2016;388(10042):376–89.
- [135].Kendler KS. What psychiatric genetics has taught us about the nature of psy-chiatric illness and what is left to learn. Mol Psychiatry 2013;18(10):1058–66.
- [136]. Maes M. Depression is an inflammatory disease, but cell-mediated immune activation is the key component of depression. Prog Neuropsychopharmacol Biol Psychiatry 2011;35(3):664–75.
- [137]. Schmitt A, Malchow B, Hasan A, Falkai P. The impact of environmental factors in severe psychiatric disorders. Front Neurosci 2014;8:19.
- [138]. Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. Nat Rev Neurosci 2012;13(10):701–12.
- [139]. Maes M, Kubera M, Leunis JC, Berk M. Increased IgA and IgM responses against gut commensals in chronic depression: further evidence for in- creased bacterial translocation or leaky gut. J Affect Disord 2012;141(1):55–62.
- [140]. Maes M, Kubera M, Leunis JC, Berk M, Geffard M, Bosmans E. In depression, bacterial translocation may drive inflammatory responses, oxidative and nitrosative stress (O&NS), and autoimmune responses directed against O&NS-damaged neoepitopes. Acta Psychiatr Scand 2013;127(5):344–54.
- [141]. Gosalbes MJ, Durbán A, Pignatelli M, Abellan JJ, Jiménez-Hernández N, Pérez-Cobas AE, et al. Metatranscriptomic approach to analyze the function- al human gut microbiota. PLoS One 2011;6(3):e17447.
- [142]. Franzosa EA, Morgan XC, Segata N, Waldron L, Reyes J, Earl AM, et al. Relating the metatranscriptome and metagenome of the human gut. Proc Natl Acad Sci USA 2014;111(22):E2329–38.
- [143]. Santiago-Rodriguez TM, Naidu M, Abeles SR, Boehm TK, Ly M, Pride DT. Transcriptome analysis of bacteriophage communities in periodontal health and disease. BMC Genomics 2015;16:549.
- [144]. Arnold JW, Roach J, Azcarate-Peril MA. Emerging technologies for gut micro- biome research. Trends Microbiol 2016;24(11):887–901.
- [145]. Connon SA, Giovannoni SJ. High-throughput methods for culturing microor- ganisms in very-low-nutrient media yield diverse new marine isolates. Appl Environ Microbiol 2002;68(8):3878–85.
- [146]. Nichols D, Cahoon N, Trakhtenberg EM, Pham L, Mehta A, Belanger A, et al. Use of ichip for high-throughput *in situ* cultivation of "uncultivable" micro- bial species. Appl Environ Microbiol 2010;76(8):2445–50.
- [147]. Jung D, Seo EY, Epstein SS, Joung Y, Han J, Parfenova VV, et al. Application of a new cultivation technology, I-tip, for studying microbial diversity in fresh- water sponges of Lake Baikal, Russia. FEMS Microbiol Ecol 2014;90(2):417–23.
- [148]. Possemiers S, Verthé K, Uyttendaele S, Verstraete W. PCR-DGGE-based quantification of stability of the microbial community in a simulator of the hu-man intestinal microbial ecosystem. FEMS Microbiol Ecol 2004;49(3):495–507.
- [149]. Petrof EO, Khoruts A. From stool transplants to next-generation microbiota therapeutics. Gastroenterology 2014;146(6):1573–82.
- [150]. McDonald JA, Fuentes S, Schroeter K, Heikamp-deJong I, Khursigara CM, de Vos WM, et



- al. Simulating distal gut mucosal and luminal communities using packed-column biofilm reactors and an *in vitro* chemostat model. J Microbiol Methods 2015;108:36–44.
- [151]. Wang BL, Ghaderi A, Zhou H, Agresti J, Weitz DA, Fink GR, et al. Microfluidic high-throughput culturing of single cells for selection based on extracellular metabolite production or consumption. Nat Biotechnol 2014;32(5):473–8.
- [152]. Kim HJ, Huh D, Hamilton G, Ingber DE. Human gut-on-a-chip inhabited by microbial flora that experiences intestinal peristalsis-like motions and flow. Lab Chip 2012;12(12):2165–74.
- [153]. Rusconi R, Garren M, Stocker R. Microfluidics expanding the frontiers of mi- crobial ecology. Annu Rev Biophys 2014;43:65–91.
- [154]. Englert DL, Manson MD, Jayaraman A. Investigation of bacterial chemotaxis in flow-based microfluidic devices. Nat Protoc 2010;5(5):864–72.
- [155]. Wang Y, Ahmad AA, Sims CE, Magness ST, Allbritton NL. *In vitro* generation of colonic epithelium from primary cells guided by microstructures. Lab Chip 2014;14(9):1622–31.
- [156]. Gracz AD, Williamson IA, Roche KC, Johnston MJ, Wang F, Wang Y, et al. A high-throughput platform for stem cell niche co-cultures and downstream gene expression analysis. Nat Cell Biol 2015;17(3):340–9.
- [157]. Forbester JL, Goulding D, Vallier L, Hannan N, Hale C, Pickard D, et al. In-teraction of *Salmonella enterica* serovar Typhimurium with intestinal organoids derived from human induced pluripotent stem cells. Infect Immun 2015;83(7):2926–34.
- [158]. Leslie JL, Huang S, Opp JS, Nagy MS, Kobayashi M, Young VB, et al. Persistence and toxin production by *Clostridium difficile* within human intestinal organoids result in disruption of epithelial paracellular barrier function. Infect Immun 2015;83(1):138–45.
- [159]. Sommer MO. Advancing gut microbiome research using cultivation. Curr Opin Microbiol 2015;27:127–32.
- [160]. Dao MC, Everard A, Aron-Wisnewsky J, Sokolovska N, Prifti E, Verger EO, et al.; MICRO-Obes Consortium. *Akkermansia muciniphila* and improved met-abolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. Gut 2016;65(3):426–36.
- [161]. Chu J, Vila-Farres X, Inoyama D, Ternei M, Cohen LJ, Gordon EA, et al. Discovery of MRSA active antibiotics using primary sequence from the human microbiome. Nat Chem Biol 2016;12(12):1004–6.
- [162]. Wang J, Jia H. Metagenome-wide association studies: fine-mining the micro- biome. Nat Rev Microbiol 2016;14(8):508–22.
- [163]. Lu H, Wu Z, Xu W, Yang J, Chen Y, Li L. Intestinal microbiota was assessed in cirrhotic patients with hepatitis B virus infection. Intestinal microbiota of HBV cirrhotic patients. Microb Ecol 2011;61(3):693–703.
- [164]. Lu H, Zhang C, Qian G, Hu X, Zhang H, Chen C, et al. An analysis of microbiota- targeted therapies in patients with avian influenza virus subtype H7N9 in- fection. BMC Infect Dis 2014;14:359.
- [165]. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. N Engl J Med 2013;368(5):407–15.
- [166]. Dhiman RK, Rana B, Agrawal S, Garg A, Chopra M, Thumburu KK, et al. Probiotic VSL#3 reduces liver disease severity and hospitalization in pa- tients with cirrhosis: a randomized, controlled trial. Gastroenterology 2014;147(6):1327–37.e3.
- [167]. Lv LX, Hu XJ, Qian GR, Zhang H, Lu HF, Zheng BW, et al. Administration of *Lactobacillus salivarius* LI01 or *Pediococcus pentosaceus* LI05 improves acute liver injury induced by *D*-galactosamine in rats. Appl Microbiol Biotechnol 2014;98(12):5619–32.
- [168]. Atarashi K, Tanoue T, Oshima K, Suda W, Nagano Y, Nishikawa H, et al. Treg induction by a rationally selected mixture of Clostridia strains from the hu- man microbiota. Nature 2013;500(7461):232-6.
- [169]. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science 2013;342(6161):967–70.
- [170]. Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Hannani D, et al. The intestinal microbiota modulates the anticancer immune effects of cyclo-phosphamide. Science 2013;342(6161):971–6.
- [171]. West CE, Jenmalm MC, Kozyrskyj AL, Prescott SL. Probiotics for treatment and primary prevention of allergic diseases and asthma: looking back and moving forward. Expert Rev Clin Immunol 2016;12(6):625–39.